# ARCHIVESOFENVIRONMENTALPROTECTIONvol. 37no. 2pp. 109 - 1212011

PL ISSN 0324-8461

© Copyright by Institute of Envionmental Engineering of the Polish Academy of Sciences, Zabrze, Poland 2011

# THE EFFECT OF SOIL CONTAMINATION LEVEL AND PLANT ORIGIN ON CONTENTS OF ARSENIC, CADMIUM, ZINC, AND ARSENIC COMPOUNDS IN *MENTHA AQUATICA* L.

# JIŘINA SZÁKOVÁ,<sup>1</sup> PAVEL TLUSTOŠ,<sup>1</sup> WALTER GOESSLER,<sup>2</sup> TOMÁŠ POKORNÝ,<sup>1</sup> SILVIA FINDENIG,<sup>2</sup> JIŘÍ BALÍK<sup>1</sup>

<sup>1</sup>Department of Agroenvironmental Chemistry and Plant Nutrition, Czech University of Life Sciences, CZ-165 21 Prague 6, Czech Republic

\*Corresponding author's e-mail: szakova@af.czu.cz

<sup>2</sup>Institute of Chemistry, Analytical Chemistry, Karl-Franzens-University Graz, Universitaetsplatz 1, A-8010 Graz, Austria

Keywords: Mentha aquatica, arsenic, cadmium, zinc, speciation, contaminated area, phytostabilization.

**Abstract:** Water mint (*Mentha aquatica* L.) belongs to the arsenic tolerant plant species suitable for cultivation in Central European climate conditions. Therefore, its possible application for remediation of contaminated soil was investigated in pot and field experiments. Two *M. aquatica* plants of different origin, i) commercially market-available mint plants, and ii) plants habituated at the arsenic contaminated former mining area in southern Tuscany (Italy) were tested for their arsenic uptake, transformation, and speciation. The total arsenic concentrations in the experimental soils varied from 21 to 1573 mg As kg<sup>-1</sup>, the mobile fractions did not exceed 2% of total soil arsenic. The mint plants originating from the contaminated area were able to remove ~400  $\mu$ g of arsenic per pot, whereas the commercial plant removed a significantly lower amount (~300  $\mu$ g of arsenic per pot). Only arsenite and arsenate, but no organoarsenic compounds were identified in both stems and leaves. Arsenate was the predominant arsenic compound and reached up to 80% regardless of the origin of the mint plants. Although *M. aquatica* seems to be able to grow in contaminated soils without symptoms of phytotoxicity, its efficiency to remove arsenic from the soil is limited as can be demonstrated by total elimination of As from individual pots not exceeding 0.1%. Moreover, the application of plants originating from the contaminated site did not result in sufficient increase of potential phytoextraction efficiency of *M. aquatica*. Although not suitable for phytoextraction the *M. aquatica* plants can be used as vegetation cover of the contaminated soil at the former mining areas.

# INTRODUCTION

Surrounding the city Kutná Hora, Central Bohemia, the area of medieval silver mining, is characterized by high level of soil contamination with risk elements (especially As, Cd, and Zn). The available analytical data [1] confirmed high arsenic contents of As in soils ranging between 50 and 330 mg·kg<sup>-1</sup>. Therefore, possible remediation of these soils must be taken into account and phytoremediation suggests itself a relatively low cost and environmental friendly method leading to decrease of soil contamination level.

However, the spectrum of plant species suitable for phytoextraction of arsenic is relatively limited. Ma *et al.* [2] described the fern species *Pteris vittata* L. as capable of accumulating up to 23000 mg kg<sup>-1</sup> of As in aboveground biomass, most of which was pre-

sent as inorganic As (47–80%). Thereafter, more fern species were described as As hyperaccumulators such as *Pityrogramma calomelanos* [3], and *Pteris cretica*, *Pteris longifolia* and *Pteris umbrosa* [4]. The results suggested that *P. calomelanos* is able to release 2% of the total As from soil. However, these plants are not able to survive in Central European climatic conditions and their applicability is negligible.

*Mentha aquatica* L. (*Lamiaceae*) was described as the plant characterized by a high content of essential oil, phenolics, and flavanoids resulting in anti-inflammatory, antioxidant, and antibacterial action of this traditional medicinal plant [5–7]. Moreover, *Mentha spp.* is frequently discussed among the other aquatic macrophytes as the potential bioindicator of risk element pollution of water [8] and soil [9].

Zurayk *et al.* [10, 11] compared twelve Mediterranean plant species characterized as hydrophytes and found that only *M. aquatica* was able to accumulate sufficient amounts of Cd, Cr, and Ni and it was considered as a bioindicator for these elements. Growth rates remained high during the experimental period, indicating that the plants were little affected by the presence of metal at the experimental concentration. In the case of lead, low root-leaf transport was observed resulting in low lead accumulation in aboveground biomass [12]. The plants are usually able to change their tolerance to risk elements according to the environmental conditions. The populations of plants growing in contaminated soil are able to adapt their tolerance to actual metal contents in soil [13]. *M. aquatica* plants sampled at an arsenic contaminated site (total content of arsenic in soil was 266 mg·kg-1) demonstrated arsenic content in aboveground biomass up to 216 mg·kg-1 and in roots up to 540 mg·kg-1 [14] without apparent symptoms of arsenic phytotoxicity. Therefore, *M. aquatica* seems to be a promising plant to be tested for potential phytoremediation of the arsenic contaminated soils.

In our previous experiments [15] the mint plants tested in rhizobox experiments did not reach the arsenic contents presented by Baroni *et al.* [14], although both total and mobile arsenic contents in soil were comparable to the levels documented in the mentioned paper. Moreover, the dominant portion of arsenic retained in roots limits the potential use of this plant for phytoremediation technologies. To verify the findings ascertained in model conditions, field experiments under real conditions of the contaminated site are necessary.

The main objectives of our pot and field experiments were i) to compare the growth potential and arsenic uptake by commercially available plants of *M. aquatica* and arsenic-tolerant plants originating from the arsenic contaminated site; ii) to compare the arsenic compound transformations within aboveground biomass of both experimental plants; and iii) to assess potential phytoremediation ability of both plants in field conditions.

# MATERIAL AND METHODS

#### Pot experiment

The area of Mokrsko (located 35 km south of Prague) has a high geogenic As content due to gold arsenopyrite occurrence. Kutná Hora (70 km east of Prague) soils are contaminated with arsenic, cadmium, and zinc mainly due to tailings of silver mining in the Middle Ages and furthermore the concentrations of Cd and Zn were always substantially higher [1]. The sampling areas were selected at the suburb of the Malín municipality close to a medieval silver mine (so-called gallery of 14 assistants) close to a small streamlet named

Beránka. The location was described and mapped elsewhere [16]. Slightly contaminated Fluvisols from Píšťany (80 km north-west from Prague) was added to the set of experimental soils. Total element contents and main soil characteristics are summarized in Table 1, the available contents of macro- and micronutrients according to Mehlich III extraction procedure [17] are given in Table 2.

Table 1. Total element contents and basic characteristics of experimental soils; data are presented as mean  $\pm$  standard deviation, n = 3

Soil	As mg kg <sup>-1</sup>	Cd mg kg <sup>-1</sup>	Pb mg kg <sup>-1</sup>	Zn mg kg <sup>-1</sup>	рН	TOC %	CEC mmol kg <sup>-1</sup>
Kutná Hora meadow	$1573 \pm 46$	$4.9\pm0.03$	$30.0 \pm 0.5$	$499\pm61$	$6.97 \pm 0.03$	$4.02 \pm 0.25$	$148 \pm 12$
Kutná Hora arable	$1312 \pm 103$	$19.5 \pm 0.4$	$79.0 \pm 0.1$	1931 ± 56	$7.41 \pm 0.03$	$0.95\pm0.00$	$380 \pm 15$
Mokrsko	$839 \pm 112$	$0.03 \pm 0.01$	$11.0\pm0.5$	$238 \pm 2$	$5.36 \pm 0.04$	$0.78\pm0.02$	$116 \pm 8$
Píšťany	$21 \pm 1$	$0.83\pm0.06$	$46 \pm 9$	$100 \pm 4$	$6.87\pm0.02$	$1.50\pm0.03$	$127 \pm 5$

TOC – total organic carbon

CEC – cation-exchange capacity

Table 2. Available contents of macro-and micronutrients in soils determined by Mehlich III extraction procedure [17]; data are presented as mean $\pm$ standard deviation, n = 3

Cail	Р	K	Ca	Mg	Fe	Mn	В
5011	mg.kg <sup>-1</sup>						
Kutná Hora	$1.10 \pm 0.12$	$103 \pm 7$	$14060 \pm 659$	$201 \pm 6$	$191 \pm 4$	$30.8 \pm 2.2$	$4.17 \pm 1.08$
meadow					-		
Kutná Hora arable	56.2 ± 1.1	610 ± 11	7210 ± 224	$543 \pm 8$	$164 \pm 9$	32.7 ± 1.0	$7.88 \pm 0.20$
Mokrsko	$33.8 \pm 1.1$	$90.7 \pm 16.9$	$1715 \pm 24$	$196\pm12$	$230\pm1$	$57.1 \pm 0.0$	$3.29 \pm 1.06$
Píšťany	$123 \pm 2$	$173 \pm 9$	$3091 \pm 1$	$220 \pm 5$	$372 \pm 3$	$97.4\pm3.3$	$1.99\pm0.38$

Before planting, soil samples were collected from individual pots, air dried at 20°C, ground in a mortar and passed through a 2-mm plastic sieve [18] and analyzed for its total and mobile element content. Plants were cultivated in 6-litre plastic pots with 5 kg of airdry soil and four replicates were used for each soil. NPK (0.5 g N, 0.16 g P, 0.4 g K per pot as inorganic salts solutions) was added before sowing and *M. aquatica* was cultivated for two vegetation periods. The *M. aquatica* plants were of different origin, i) commercially market-available mint plants and ii) plants habituated at the arsenic contaminated former mining area in southern Tuscany (Italy) purchased by the University of Siena, Dipartimento Sci. Ambientali, Sez. Geochim. Ambientale, Siena, Italy. The plants were prepared by vegetative propagation from one plant cultivated in uncontaminated soil. Soil moisture was regularly controlled and kept at 60% of the maximum water holding capacity (MWHC). The aboveground biomass was harvested at the end of the vegetation period, gently washed by deionized water, checked for fresh biomass, freeze-dried (Lyovac GT-2, Germany), ground and analyzed.

# Field experiment

The same treatments as in the pot experiment were performed in a precise field experiment (plot size: 4 m<sup>2</sup>) at the locations of Kutná Hora arable and Kutná Hora meadow for three vegetation periods. Twenty five plants of the commercial mint in 5 rows and 12 plants of the mint originating from the contaminated site in 3 rows were planted in each plot. The plants were planted under natural conditions, only weed plants were regularly removed. No fertilizers and/or pesticides were applied for the entire duration of the experiment. The aboveground biomass was harvested at the end of each vegetation period, gently washed by deionised water, checked for fresh biomass, freeze-dried (Lyovac GT-2, Germany), ground and analyzed.

#### Analytical methods

Total element concentrations in soil were determined in digests obtained by two-step decomposition as follows: 0.5 g of sample was decomposed by dry ashing in a mixture of oxidizing gases  $(O_2 + O_3 + NO_x)$  in an Apion Dry Mode Mineralizer (Tessek, CZ) at 400°C for 10 h; the ash was then dissolved in a mixture of HNO<sub>3</sub> + HF, evaporated to dryness at 160°C and dissolved in diluted Aqua Regia [19]. Certified reference material RM 7001 Light Sandy Soil (Analytika, CZ) containing 12.3 ± 1.1 mg As kg<sup>-1</sup>, 0.32 ± 0.05 mg Cd kg<sup>-1</sup>, 43.8 ± 3.7 mg Pb kg<sup>-1</sup>, and 120 ± 7 mg Zn kg<sup>-1</sup> was used for quality assurance of the analytical data of total element determination, and 12.8 ± 1.5 mg As kg<sup>-1</sup>, 0.30 ± 0.01 mg Cd kg<sup>-1</sup>, 39.9 ± 1.3 mg Pb kg<sup>-1</sup>, 113±6 mg Zn kg<sup>-1</sup> was determined in this sample.

For determination of mobile element portions, aliquots of the soil samples were extracted with 0.05 mol L<sup>-1</sup> aqueous  $(NH_4)_2SO_4$  solution at a solid/liquid ratio of 1/25 (1 g + 25 ml) for 4 hours [20]. Each extraction was done three times. All the chemicals used were of electronic grade purity and were purchased from Analytika and Lach-Ner Ltd., Czech Republic. For the centrifugation of the extracts, the Hettich Universal 30 RF (Germany) device was used. The pH was determined using deionised water or 0.2 M KCl (w/v = 1 + 2.5). Cation-exchange capacity (CEC) was calculated as the sum of Ca, Mg, K, and Al extractable in 0.1 M BaCl<sub>2</sub> (w/v = 1+20 for 2 hours) [21]. Total organic carbon (TOC) was determined spectrophotometrically after the oxidation of organic matter by K<sub>2</sub>Cr<sub>2</sub>O<sub>7</sub> [22]. The available contents of nutrients were determined by the Mehlich III soil extraction procedure [17] using flame atomic absorption spectroscopy (FAAS, VARIAN SpectrAA-300, Australia) (for Ca, K and Mg) and ICP-OES with axial plasma configuration (Varian VistaPro, Varian, Australia, for P, Fe, B, Mn), see Table 2.

Plant samples were decomposed using the dry ashing procedure as follows: An aliquot (~1 g) of the dried and powdered aboveground biomass was weighed to 1 mg into a borosilicate glass test-tube and decomposed in a mixture of oxidizing gases ( $O_2 + O_3 + NO_x$ ) at 400°C for 10 hours in Dry Mode Mineralizer Apion (Tessek, Czech Republic). The ash was dissolved in 20 mL of 1.5% HNO<sub>3</sub> (electronic grade purity, Analytika Ltd., Czech Republic) and kept in glass tubes until the analysis [23]. Aliquots of the certified reference material RM NCS DC 73350 Poplar leaves (purchased from Analytika, CZ) were mineralized under the same conditions for quality assurance of the total arsenic contents in experimental plants. This material was certified to contain the following: 0.37  $\pm$  0.06 mg As kg<sup>-1</sup>, 0.32  $\pm$  0.07 mg Cd kg<sup>-1</sup>, 1.5  $\pm$  0.3 mg Pb kg<sup>-1</sup>, and 37  $\pm$  3 mg Zn kg<sup>-1</sup> was determined 0.36  $\pm$  0.02 mg As kg<sup>-1</sup>, 0.34  $\pm$  0.05 mg Cd kg<sup>-1</sup>, 1.6  $\pm$  0.2 mg Pb kg<sup>-1</sup>, and 35  $\pm$  3 mg Zn kg<sup>-1</sup>. For determination of arsenic compounds in *M. aquatica* biomass aliquots (~500 mg) of the dried and powdered aboveground biomass or roots were weighed to 0.1 mg into 10-mL screw-capped polyethylene tubes and 10 mL of 20 mmol L<sup>-1</sup> ammonium phosphate buffer was added [24]. The closed tubes were fastened to the arms of a cross-shaped rotor and turned top over bottom at 45 rpm for 14 hours. The mixtures were then centrifuged for 10 min. at 3000 rpm, and filtered through a 0.22  $\mu$ m cellulose-nitrate ester filter (Millex-GS, Milipore, Bedford, MA, USA). Aliquots of this solution (20  $\mu$ l) were chromatographed.

The total arsenic concentrations in the plant biomass decomposed by dry ashing procedure and in soil digests, soil, and plant extracts were determined by hydride generation atomic absorption spectrometry (Varian AA280Z, Varian, Australia), equipped with a continuous hydride generator VGA-77 [25] where a mixture of potassium iodide and ascorbic acid was used for pre-reduction of the sample and the extract was acidified by HCl before measurement. High arsenic concentrations as well as cadmium, and zinc were determined by ICP-OES with axial plasma configuration (Varian VistaPro, Varian, Australia).

An Agilent 1100 solvent delivery unit (Germany) together with a Hamilton PRP-X100 (USA) anion-exchange column (250 mm x 4.1 mm i.d., spherical 10- $\mu$ m particles of a styrene-divinylbenzene copolymer with trimethylammonium exchange sites) was used for the separation of As(III), dimethylarsinic acid (DMA), methylarsonic acid (MA), and As(V). An aqueous 0.02 mol L<sup>-1</sup> NH<sub>4</sub>H<sub>2</sub>PO<sub>4</sub> solution pH 6.0 at a flow rate of 1.5 mL min<sup>-1</sup> served as mobile phase. The column effluent was introduced into the plasma of the ICPMS (Agilent 7500ce) for arsenic selective-detection [26].

## RESULTS AND DISCUSSION

#### Pot experiment

Table 1 documents a high level of soil arsenic contamination in the Kutná Hora location reaching  $1573 \pm 46$  mg As kg<sup>-1</sup> as also reported by other authors [27]. According to the public notice released by Czech Ministry of the Environment [18] the element concentrations in soils cannot exceed a maximum of 30 mg kg<sup>-1</sup> As, 1.0 mg kg<sup>-1</sup> Cd, 140 mg kg<sup>-1</sup> Pb and 200 mg kg<sup>-1</sup> Zn. The total arsenic and zinc concentrations reached the legislation threshold limit in all the investigated soils with the exception of the soil Píšťany. Elevated concentrations of cadmium were determined in both soils from the Kutná Hora location. However, mobile portions of arsenic in the soils are relatively low. The water extractable arsenic portions in the soils of Kutná Hora characterized by high total organic matter content and/or high sorption capacity do not exceed 0.3% of total arsenic content in soil. On the contrary, the mobile content of arsenic in the sandy soil Píšťany reached 2% of total arsenic content, most probably due to a low level of CEC. No effect of changing pH levels was observed although a decreasing As mobility with increasing sorption capacity of the soil occurred in our previous experiment [28]. The arsenic mobility within the set of experimental soil is usually not related significantly to some of the individual soil characteristics but is affected by a whole complex of soil parameters [19, 29]. Therefore, there is a big problem in finding the soil parameter unambiguously responsible for changing As mobility in individual soils. For cadmium, the mobile contents varied between 0.5% (Kutná Hora) and 9% (Mokrsko) of the total contents. In the case of Zn, the mobile contents varied from 0.2% (Píšťany) to 8% (Mokrsko).

Comparing the yield of aboveground plant biomass (Table 3) a higher mass of leaves vs. stems was reported where no effect of soil contamination level was reported.

Table 3. Total yield of aboveground plant biomass per pot (g); data are presented as mean  $\pm$  standard deviation, n = 4; The averages marked by the same letter did not significantly differ at  $\alpha$  = 0.05 within individual columns

1 <sup>st</sup> vegetation period									
Soil	commerc	ial plants	plants from co	ntaminated site					
5011	leaves	stems	leaves	stems					
Kutná Hora meadow	8.2 ± 1.9 <sup>b</sup>	$6.5\pm2.7$ ab	13.6 ± 1.2 °	$8.4 \pm 1.9$ bcd					
Kutná Hora arable	12.6 ± 1.0 °	$10.4 \pm 1.0$ <sup>cd</sup>	$16.5 \pm 1.9$ <sup>d</sup>	$11.1 \pm 1.8$ <sup>d</sup>					
Mokrsko	$5.8 \pm 1.5^{ab}$	7.4 ± 1.5 <sup>b</sup>	$4.5 \pm 1.7$ <sup>a</sup>	$3.88 \pm 0.30$ a					
Píšťany	11.9 ± 1.3 °	$8.1 \pm 1.7 \text{ bc} \qquad 14.00 \pm 2.4 \text{ cd}$		$8.6 \pm 1.9$ bcd					
2 <sup>nd</sup> vegetation period									
Coil.	commerc	ial plants	plants from contaminated site						
5011	leaves	stems	leaves	stems					
Kutná Hora meadow	$3.85 \pm 0.50$ a	$0.95 \pm 0.11$ a	$10.3 \pm 2.2$ <sup>cd</sup>	$4.1 \pm 1.2$ bc					
Kutná Hora arable	$7.8 \pm 2.5$ bc	$6.7\pm2.6$ <sup>cd</sup>	11. $8 \pm 0.2$ <sup>d</sup>	$8.8 \pm 1.5$ d					
Mokrsko	$3.7 \pm 1.2$ a	$3.5 \pm 1.1^{ab}$	$5.2 \pm 1.9$ <sup>ab</sup>	$4.6 \pm 2.0$ bc					
Píšťany	$6.7 \pm 2.8$ abc	$3.38\pm0.95~^{ab}$	19.5 ± 3.5 °	$8.1 \pm 1.7$ <sup>d</sup>					

In most cases, a higher yield of aboveground biomass was produced by the *M. aquatica* plants originating from the contaminated site. The highest total arsenic contents (Table 4) were determined in the soils with lower sorption capacity (Kutná Hora meadow, Mokrsko) compared to Kutná Hora arable with a CEC level of  $380 \pm 15$  mmol.kg<sup>-1</sup> confirming that the increasing element content in plants is not affected solely by the increasing total element content in soil [30].

Table 4. Total arsenic contents in aboveground plant biomass – pot experiment (mg.kg<sup>-1</sup>); data are presented as mean  $\pm$  standard deviation, n = 4. The averages marked by the same letter did not significantly differ at  $\alpha = 0.05$  within individual columns

1 <sup>st</sup> vegetation period									
Coil	commerc	cial plants	plants from co	ntaminated site					
5011	leaves	stems	leaves	stems					
Kutná Hora meadow	$13.7 \pm 5.0$ <sup>d</sup>	3.65 ± 1.42 b	$10.2 \pm 4.2$ <sup>cd</sup>	$2.44\pm0.96~^{ab}$					
Kutná Hora arable	$5.03 \pm 0.38$ <sup>ab</sup>	$1.60 \pm 0.53$ <sup>ab</sup>	$8.13 \pm 2.43$ bc	$1.49 \pm 0.46$ a					
Mokrsko	$10.0\pm0.9$ <sup>cd</sup>	$1.94 \pm 0.31$ ab	$12.4 \pm 4.5$ <sup>cd</sup>	7.30 ± 3.01 °					
Píšťany	$1.28 \pm 0.51$ a	$1.12 \pm 0.43$ a	$1.71 \pm 0.83$ a	$0.75 \pm 0.59$ a					
2 <sup>nd</sup> vegetation period									
Coil	commerc	cial plants	plants from co	ntaminated site					
5011	leaves	stems	leaves	stems					
Kutná Hora meadow	$42.2 \pm 6.8$ <sup>d</sup>	$13.8 \pm 5.6$ <sup>a</sup>	20.24 ± 5.58 °	8.85 ± 1.33 ª					
Kutná Hora arable	$7.20 \pm 1.03$ <sup>ab</sup>	$2.98 \pm 0.93$ a	$4.60 \pm 0.98$ a	$1.77 \pm 0.12$ a					
Mokrsko	37.7 ± 12.3 °	41.4± 11.4 <sup>b</sup>	$17.1 \pm 7.7$ bc	3.15 ± 2.35 ª					
Píšťany	$4.46 \pm 2.21$ a	$3.50 \pm 2.313$ a	$4.39 \pm 1.29$ <sup>a</sup>	$2.27 \pm 0.35$ a					

The effect of P on the increasing mobility of arsenate due to a competitive sorption of arsenate and phosphate in soils was highlighted by many authors [31, 32]. In our case, however, the plants were fertilized and the potential effect of different P levels in soils was suppressed. Apparently, leaves are able to accumulate arsenic more intensively than stems, and better rooting of plants in the second vegetation period resulted in better arsenic uptake, especially in the case of commercial plants. Due to a higher yield of plant biomass originating from the contaminated site, the total arsenic uptake per pot tended to higher values in these samples (Figure 1).



Kutná Hora Kutná Hora Mokrsko Píšťany meadow arable

Figure 1. Total uptake of arsenic by aboveground biomass of M. aquatica – pot experiment; A) commercial plants, B) plants from the contaminated site

Chaney [33] argued that plant arsenic levels between 3 and 10 mg kg<sup>-1</sup> can be considered as phytotoxic. Our experimental plants demonstrated no visible symptoms of phytotoxicity although the arsenic contents in mint leaves exceeded these levels even 4-fold in some cases. According to Directive No. 2002/32/ES [34] the maximum values of elements in raw feedstuffs (we calculated 12% of humidity) are 2 mg kg<sup>-1</sup> of arsenic. Therefore, the potential transfer of the elements originating from the *M. aquatica* plants growing in the contaminated soil to the food chain by means of herbivorous animals should be taken into account. Zheljazkov et al. [35] found no detectable amount of Cd, Cu, or Pb in the essential oils of peppermint, basil, and dill and supported the use of aromatic plants as alternative crops for Cd, Pb, and Cu enriched soils. The potential effect of previous adaptation of a particular water mint sample collected directly at the extremely contaminated site was not observed. The highest As concentration in plants was described in submerged roots and stems, an order of magnitude higher than the aerial stems and leaves due to physicochemical adsorption, most probably due to co-precipitation with Fe at the submerged parts of stems [36, 37]. In our case, the plants were planted in the soils where the humidity was kept at 60% of MWHC. This was not representative of typical wetlands with submerged parts of the plants and presence of "iron plaque" at plant stems cannot be expected.

The results of determination of As compounds in mint plants showed dominating pools of arsenate in both stems and leaves of *M. aquatica* plants (Table 5) followed by arsenite and small percentages of DMA (not exceeding 1.5%) and occasionally MA.

In our previous rhizobox experiment only arsenite and arsenate (no organoAs) compounds were identified in both aboveground biomass and roots [15]. In this pot experiment the presence of organoarsenicals can be explained by a longer duration of the experiment resulting in a higher uptake of arsenic compared to the rhizobox experiment. A small percentage of organic compounds of As were determined, for example, in stems and leaves of pepper plants [24] and described by other authors in many different plant species [38, 39]. No significant differences occurred between commercial plants and plants originating from the contaminated site.

For potential plant use in phytoextraction of risk element tolerance of plants to the high soil contamination level, high yields of aboveground biomass and predominant accumulation of the elements in aboveground plant biomass are necessary [40]. In our case, during two vegetation periods, the *M. aquatica* plants were able to remove only 393 µg As per pot in the case of the plants originating from the contaminated site and 299 µg As per pot for the commercial plants. As apparent from Figure 1, the plants tended in most cases to a higher total uptake of arsenic in the second vegetation period suggesting gentle adaptation of the plants to the cultivation conditions. The effective removal of the risk element from the soil is usually evaluated using transfer factor values given as a ratio of element concentration in the aboveground biomass of plant and in the soil. In this context, Iskandar and Kirkham [41] found the levels of the transfer factors of risk elements decreasing in order Cd > Zn > Mn > Cu > Co > Pb > Cr > As. In our experiment the transfer factors varied between 0.006 and 0.379 for the commercial plants and between 0.006 and 0.317 for the plants originating from the contaminated site confirming a limited ability of mint plants to accumulate arsenic in the aboveground biomass. Moreover, our previous rhizobox experiment [24] showed that a significant portion of the element (60% at the highly contaminated soil) was retained in roots. Therefore, the potential effectivity

			- ( J J	)	5	_			
Stems	As(III)		DMA		MA		As(V)		sum
commercial plants	mg.kg <sup>-1</sup>	% of sum	mg.kg <sup>-1</sup>						
Kutná Hora meadow	$1.69 \pm 0.01$	33.0	$0.034 \pm 0.00$	0.7	$0.016 \pm 0.01$	0.31	$3.39\pm0.00$	66.1	$5.14 \pm 0.01$
Kutná Hora arable	$0.229 \pm 0.03$	13.9	$0.024 \pm 0.00$	1.4	$0.008\pm0.00$	0.46	$1.40 \pm 0.11$	84.2	$1.66\pm0.10$
Mokrsko	$6.1 \pm 1.08$	8.2	$0.215 \pm 0.03$	0.3	< 0.001	ı	$67.0 \pm 7.70$	91.5	$73.4 \pm 8.72$
Píšťany	$0.347 \pm 0.21$	24.8	$0.00 \pm 0.00$	0.7	< 0.001	ı	$0.95\pm0.09$	74.7	$1.31 \pm 0.29$
plants from contaminated site									
Kutná Hora meadow	$1.70 \pm 0.30$	29.3	$0.049 \pm 0.00$	0.8	$0.005\pm0.00$	0.09	$4.13 \pm 0.79$	69.8	$5.88 \pm 0.64$
Kutná Hora arable	$0.200\pm0.00$	17.5	$0.012 \pm 0.00$	1.1	$0.001\pm0.00$	0.06	$0.93\pm0.09$	81.4	$1.15 \pm 0.09$
Mokrsko	$1.85 \pm 0.77$	15.1	$0.060 \pm 0.02$	0.5	$0.004\pm0.00$	0.03	$10.7 \pm 2.48$	84.4	$12.6 \pm 2.23$
Píšťany	$0.126 \pm 0.01$	14.8	$0.007 \pm 0.00$	0.8	< 0.001	ı	$0.73 \pm 0.15$	84.4	$0.868\pm0.15$
Leaves	As(III)		DMA		MA		As(V)		sum
commercial plants	mg/kg	% of sum	mg/kg	% of sum	ug/kg	% of sum	Mg/kg	% of sum	mg/kg
Kutná Hora meadow	$8.74 \pm 1.01$	46.1	$0.170 \pm 0.06$	0.90	$0.057 \pm 0.05$	0.30	$9.98\pm0.62$	52.7	$18.9 \pm 0.89$
Kutná Hora arable	$0.946\pm0.08$	28.9	$0.027 \pm 0.00$	0.83	< 0.001	ı	$2.30\pm0.19$	70.2	$3.28 \pm 1.42$
Mokrsko	$8.03 \pm 1.23$	29.8	$0.307 \pm 0.02$	1.14	$0.018 \pm 0.01$	0.06	$18.6 \pm 1.6$	69.0	$27.0 \pm 1.53$
Píšťany	$0.175 \pm 0.02$	17.7	$0.006 \pm 0.00$	0.58	< 0.001	ı	$0.81\pm0.06$	81.7	$0.989 \pm 0.07$
plants from contaminated site									
Kutná Hora meadow	$5.20 \pm 0.03$	47.1	$0.085 \pm 0.02$	0.77	$0.008 \pm 0.002$	0.1	$5.76 \pm 0.61$	52.0	$11.1 \pm 0.54$
Kutná Hora arable	$0.972 \pm 0.04$	42.0	$0.028\pm0.00$	1.21	< 0.001		$1.31\pm0.08$	56.8	$2.3 \pm 0.04$
Mokrsko	$1.51 \pm 0.28$	21.7	$0.104\pm0.03$	1.46	< 0.001		$5.53 \pm 1.78$	76.8	$7.1 \pm 2.06$
Píšťany	$0.606 \pm 0.16$	23.2	$0.014\pm0.00$	0.53	< 0.001		$1.97 \pm 0.11$	76.2	$2.6 \pm 0.23$

Table 5. Arsenic compounds in above round plant biomass – pot experiment, average of both vegetation periods; data are presented as mean $\pm$ standard deviation, n = 3

of the *M. aquatica* plants for arsenic phytoextraction is negligible but the plants showed high tolerance to arsenic contamination of soil regardless of plant origin and soil arsenic level. Similarly, *Agrostis tenuis* plants growing without phytotoxicity symptoms in the soil containing up to 26500 mg kg<sup>-1</sup> As were able to accumulate 3470 mg kg<sup>-1</sup> As [42] confirming a high tolerance of these plants to elevated arsenic contents in soil without their hyperaccumulation parameters.

## Field experiment

For verification of the phytoremediation potential of plants, a controlled field experiment is necessary compared with the pot experiment. Plant growth in the Kutná Hora arable soil occurred without any problems and/or any phytotoxicity symptoms. The yields of aboveground biomass of commercial plants varied between 8.1 and 13.5 g of dry matter per plant while plants originating from the contaminated site yielded between 7.0 and 16.0 g of dry matter per plant. In contrast to the pot experiment, plant growth at the Kutná Hora meadow site deteriorated and the yields varied between 2.5 and 7.6 g of dry matter per plant of plants, and between 0.6 and 7.6 g of dry matter per plant of plants originating from the contaminated site. The lower yield can be affected by different growing conditions at individual sites where the Kutná Hora meadow experimental plot was located (on a shady and sloping field).

The arsenic contents in commercial *M. aquatica* plants varied between 1.81 and 8.07 mg As kg<sup>-1</sup> of dry matter from the Kutná Hora arable soil and between 3.41 and 9.45 mg As kg<sup>-1</sup> of dry matter from the Kutná Hora meadow soil. In the case of plants originating from the contaminated site the arsenic contents varied between 3.02 and 100 mg kg<sup>-1</sup> of dry matter from the Kutná Hora meadow soil and between 3.24 and 7.08 mg kg<sup>-1</sup> of dry matter from the Kutná Hora arable soil (Table 6).

location	season	plant	As	Cd	Zn
			mg.kg <sup>-1</sup>	mg.kg <sup>-1</sup>	mg.kg <sup>-1</sup>
	1 <sup>st</sup>	commercial plants	$9.45 \pm 3.53$	$0.29\pm0.08$	$60.0 \pm 2.9$
Kutná Hora meadow		plants from contaminated site	$100 \pm 4$	$1.66\pm0.09$	$178 \pm 2$
	2 <sup>nd</sup>	commercial plants	$7.38 \pm 2.92$	$0.16\pm0.03$	$80.7 \pm 16.7$
		plants from contaminated site	$15.0 \pm 0.5$	$0.31\pm0.08$	$181 \pm 11$
	3 <sup>rd</sup>	commercial plants	$3.41 \pm 0.66$	$0.13\pm0.04$	$117 \pm 17$
		plants from contaminated site	$3.02 \pm 1.80$	$0.16\pm0.02$	$190 \pm 18$
Kutná Hora arable	1 <sup>st</sup>	commercial plants	$3.78 \pm 1.19$	$0.15 \pm 0.02$	$30.1 \pm 5.4$
		plants from contaminated site	$3.24 \pm 0.95$	$0.09\pm0.03$	$24.6 \pm 1.5$
	2 <sup>nd</sup>	commercial plants	$1.81 \pm 0.05$	$0.07\pm0.02$	$71.3 \pm 3.6$
		plants from contaminated site	$5.25 \pm 0.21$	$0.43\pm0.09$	$73.1 \pm 4.8$
	3 <sup>rd</sup>	commercial plants	$8.07 \pm 0.36$	$0.05\pm0.01$	$87.2 \pm 8.1$
		plants from contaminated site	$7.08 \pm 0.24$	$0.06 \pm 0.01$	$82.3 \pm 4.9$

Table 6. Total arsenic, cadmium, and zinc contents in aboveground plant biomass – field experiment (mg.kg<sup>-1</sup>); data are presented as mean  $\pm$  standard deviation, n = 3.

In the Kutná Hora arable soil, the arsenic concentrations increased gradually during the three vegetation periods, the highest As concentrations in the Kutná Hora meadow

soil were found in the first vegetation period, while the next seasons showed significantly lower concentrations ( $\alpha = 0.05$ ). However, a higher element concentration was associated with a lower biomass yield resulting in a comparable total removal of arsenic within individual growing seasons. Due to the elevated concentrations of Cd and Zn in the experimental soils, the contents of these elements in plant biomass was determined to assess the potential source of additional stress. In contrast to arsenic, cadmium belongs to the elements that is easily translocated to the aboveground biomass of plants [43]. In our case, however, the cadmium contents in mint aboveground biomass did not exceed 2 mg·kg<sup>-1</sup> confirming low mobility and plant-availability of cadmium with an increasing pH level of the soils [44]. Zinc concentrations in plant leaves can be classified as follows: zinc content below 10 mg·kg<sup>-1</sup> of dry matter results in symptoms of its deficit in plant tissues; zinc content between 10 and 25 mg·kg<sup>-1</sup> of dry matter is still insufficient for plants; zinc content between 26 and 150 mg kg<sup>-1</sup> represents a normal level of the element; the content between 151 and 400 mg·kg<sup>-1</sup> of dry matter indicates an enhanced level, and more than 400 mg·kg<sup>-1</sup> of dry matter represents a toxic level of zinc [45]. In our case the zinc concentrations in plants varied between 25 and 190 mg·kg<sup>-1</sup> allowing us to suggest that these values did not reach the potentially toxic levels of zinc in the aboveground biomass.

The highest total removal of arsenic from  $1 \text{ m}^2$  of the plot after three vegetation periods represented 21.5 mg. This level was reached for the plants originating from the contaminated site at Kutná Hora arable soil. Transfer factors of arsenic were lower compared to the pot experiments and did not exceed 0.1, confirming that the application of *M. aquatica* plants for phytoextraction use is unreliable. Both pot and field experiments can be compared as follows: *M. aquatica* plants demonstrated good tolerance to elevated contents of arsenic in soil regardless of plant origin and cultivation conditions. The pot experiment, under more stable and controlled cultivation conditions, expectedly showed less variation of the results than was observed in the field. Both experiments, however, clearly showed that *M. aquatica* is an unsuitable plant for effective phytoextraction of arsenic in contaminated soil.

Baroni *et al.* [14] determined arsenic contents in 64 plant species in the area of the former mine in south Tuscany (Italy), and the highest arsenic contents were determined in the leaves and roots of *M. aquatica* (as mentioned above) and in roots of *Phragmites australis*. The mint plants did not reach the arsenic contents presented by Baroni *et al.* [14] in both pot and field experiments, although both total and mobile arsenic contents in soil were comparable to the levels documented in the mentioned paper. Therefore, the growth of *M. aquatica* at the extremely contaminated site did not lead to the change in arsenic uptake by these plants compared to the plants cultivated in uncontaminated soil. Due to the lack of relevant information, there was no possibility to compare physicochemical parameters of soils at our locations with the location described by Baroni *et al.* [14], although a possible presence of "iron plaque" at the stems can be taken into account.

Although not suitable for phytoextraction, the *M. aquatica* plants demonstrated high tolerance and good growth ability in the highly contaminated soil. The plants are able to create thick and permanent vegetation cover capable of surviving more subsequent vegetation periods, and may be applied towards the stabilization of the extremely contaminated sites. Moreover, the vegetation cover is stable for several vegetation periods without intensive fertilizers and/or pesticide application. However, potential environmental risks arise from elevated plant biomass arsenic content which exceeds the maximum per-

missible limits for feedstuffs. If applied towards the stabilization of contaminated land, protection against herbivorous animals as well as suitable disposal of the contaminated biomass should be achieved.

**Acknowledgements:** The authors wish to thank MSM project No. 6046070901 of the Ministry of Education, Youth and Sports, Czech Republic for their financial support, and Mr. Brian Kavalir (Ontario, Canada) for his proofreading services. The experimental plants were kindly supplied by the University of Siena, Dipartimento Sci. Ambientali, Sez. Geochim. Ambientale, Siena, Italy.

# REFERENCES

- Malec J., P. Pauliš: Kutnohorský rudní revír a projevy zaniklé důlní a hutní činnosti na jeho území. Bull. min.- petrolog. odd. Nár. Muz. v Praze, 4-5, 84–105 (1997).
- [2] Ma L.Q., K.M. Komar, C. Tu, W.H. Zhang, Y. Cai, E.D. Kennelley: A fern that hyperaccumulates arsenic - A hardy, versatile, fast-growing plant helps to remove arsenic from contaminated soils. Nature, 409, 579 (2001).
- [3] Visoottiviseth P., K. Francesconi, W. Sridokchan: *The potential of Thai indigenous plant species for the phytoremediation of arsenic contaminated land*. Environ. Pollut., **118**, 453–461 (2002).
- [4] Zhao F.J., S.J. Dunham, S.P. McGrath: Arsenic hyperaccumulation by different fern species. New Phytologist, 156, 27–31 (2002).
- [5] Agostini F., A.C.A. dos Santos, M. Rossato, M.R. Pansera, P.L. L.A. dos Santos, Serafini, R. Molon, P. Moyna: *Essential oil yield and composition of Lamiaceae species growing in southern Brazil*. Braz. Arch. Biol. Technol. 52, 473–478 (2009).
- [6] Conforti F., S. Sosa, M. Marrelli, F. Menichini, G.A. Statti, D. Uzunov, A. Tubaro, R. Della Loggia: In vivo anti-inflammatory and in vitro antioxidant activities of Mediterranean dietary plants. J. Ethnopharmacol. 116, 144–151 (2008).
- [7] Esmaeili A., A. Rustayian, S. Masoudi, K. Nadji: Composition of the essential oils of Mentha aquatica L. and Nepeta meyeri Benth. from Iran. J. Essent. Oil Res. 18, 263–265 (2006).
- [8] Samecka-Cymerman A., A.J. Kempers: Bioindication of heavy metals with aquatic macrophytes: The case of a stream polluted with power plant sewages in Poland. J. Toxicol. Environ. Health 62, 57–67 (2000).
- [9] Zheljazkov V.D., N.E. Nielsen: *Effect of heavy metals on peppermint and cornmint*. Plant Soil. 178, 59–66 (1996).
- [10] Zurayk R., B. Sukkariyah, R. Baalbaki: Common hydrophytes as bioindicators of nickel, chromium and cadmium pollution. Water Air Soil Pollut. 127, 373–388 (2001).
- [11] Zurayk R., B. Sukkariyah, R.Baalbaki, D. Abi Ghanem: Ni phytoaccumulation in Mentha aquatica L. and Mentha silvestris L. Water Air Soil Pollut. 139, 355–364 (2002).
- [12] Saygideger S., M. Dogan: Influence of pH on lead uptake, chlorophyll and nitrogen content of Nasturtium officinale R. Br. and Mentha aquatica L. J. Environ. Biol. 26, 753–759 (2005).
- [13] Sanità di Toppi L., R. Gabbrielli: Response to cadmium in higher plants. Environ Exp Bot, 41, 105–130 (1999).
- [14] Baroni F., A. Boscagli, L.A. DiLella, G. Protano, F. Riccobono: Arsenic in soil and vegetation of contaminated areas in southern Tuscany (Italy). J. Geochem. Expl. 81, 1–14 (2004).
- [15] Száková, J; P. Tlustoš; W Goessler; S. Findenig, E. Richtrová, J. Balik: A comparison of arsenic mobility in Phaseolus vulgaris, Mentha aquatica, and Pteris cretica rhizosphere. Centr. Eur. J. Biol. 4, 107–116 (2009).
- [16] Králová L., J. Száková, Š. Kubík, P. Tlustoš, J. Balík: The variability of arsenic and other risk element uptake by individual plant species growing on contaminated soil. Soil Sedim. Contam. 19, 617–634 (2010).
- [17] Mehlich A. Mehlich: 3 Soil Test Extractant: A modification of Mehlich 2 Extractant. Comm. Soil Sci. Plant Anal. 15, 1409–1416 (1984).
- [18] Public notice No. 13/1994, regulating some details concerning the preservation of agricultural lands available. Czech Ministry of the Environment, Prague (1994).
- [19] Száková J., P. Tlustoš, J. Balík, D. Pavlíková, M. Balíková: *Efficiency of extractants to release As, Cd, and Zn from main soil compartments*, Analusis 28, 808–812 (2000).

- [20] Wenzel W.W., N. Kirchbaumer, T. Prohaska, G. Stingeder, E. Lombi, D.C. Adriano: Arsenic fractionation in soils using an improved sequential extraction procedure. Anal. Chim. Acta 436, 1–15 (2001).
- [21] ISO 11260. Standard of Soil Quality Determination of Effective cation Exchange Capacity and Base Saturation Level Using Barium Chloride Solution. International Organization for Standardization (1994).
- [22] Sims J.R., V.A. Haby: Simplified colorimetric determination of soil organic matter. Soil Sci. 112, 137– 141 (1971).
- [23] Miholová D., P. Mader, J. Száková, A. Slámová, Z. Svatoš: Czechoslovakian biological certified reference materials and their use in the analytical quality assurance system in a trace element laboratory. Fresenius J. Anal. Chem. 345, 256–260 (1993).
- [24] Száková J., P. Tlustoš, W.Goessler, D. Pavlíková, J. Balík: Comparison of mild extraction procedures for determination of arsenic compounds in different parts of pepper plants (Capsicum annum, L.). Appl. Organomet. Chem. 19, 308–314 (2005).
- [25] Brodie K., B. Frary, B. Sturman, L. Voth: An automated vapor generation accessory for atomic absorption analysis. Varian Instruments at Work AA 38, 1–8 (1983).
- [26] Schmeisser E., W. Goessler, N. Kienzl, K.A. Francesconi: Volatile analytes formed from arsenosugars: Determination by HPLC-HG-ICPMS and implications for arsenic speciation analyses. Anal. Chem. 76, 418–423 (2004).
- [27] Hyršl J., M. Kaden: Eine Paragenese von Eisen-Arsenaten von Kaňk bei Kutná Hora in Böhmen und Munzig bei Meissen in Sachsen. Aufschluss 43, 95–102 (1992).
- [28] Száková J., P. Tlustoš, D. Pavlíková, A. Hanč, M. Batysta: Effect of addition of ameliorative materials on the distribution of As, Cd, Pb, and Zn in extractable soil fractions. Chem. Pap. 61, 276–281 (2007).
- [29] Hudson-Edwards K.A., S.L. Houghton, A. Osborn: Extraction and analysis of arsenic in soils and sediments, Trends Anal. Chem. 23, 745–752 (2004).
- [30] Brooks R.R.: Plants that hyperaccumulate heavy metals. CAB INTERNATIONAL. Walligford. UK. New York. USA (1998).
- [31] Smith E., R Naidu, A.M. Alston: Chemistry of arsenic in soils: I. Sorption of arsenate and arsenite by four Australian soils. J. Environ. Qual. 28, 1719–1726 (1999).
- [32] Smith E, R. Naidu, A.M. Alston: Chemistry of arsenic in soils: II. Effect of phosphorus, sodium, and calcium on arsenic sorption. J. Environ. Qual. 31, 557–563 (2002).
- [33] Chaney R. L. In: Final Report of the Workshop on the Intern. Transportation. Pan American Health Organization, Washington (1985).
- [34] Directive No. 2002/32/ES of European Parliament and Council of Europe concerning xenobiotics in feedstuffs (2002).
- [35] Zheljazkov V.D., L.E. Craker, B.S. Xing: *Effects of Cd, Pb, and Cu on growth and essential oil contents in dill, peppermint, and basil.* Environ. Exper. Bot. 58, 9–16 (2006).
- [36] Otte M.L., C.C. Kearns, M.O. Doyle: Accumulation of arsenic and zinc in the rhizosphere of wetland plants. Bull. Environ. Contam. Toxicol. 55, 154–161 (1995).
- [37] Robinson B., N. Kim, M. Marchetti, C. Moni, L. Schroeter, C. van den Dijssel, G. Milne, B. Clothier: Arsenic hyperaccumulation by aquatic macrophytes in the Taupo Volcanic Zone, New Zealand. Environ. Exper. Bot. 58, 206–215 (2006).
- [38] Geiszinger A., W. Goessler, W. Kosmus: Organoarsenic compounds in plants and soil on top of an ore vein. Appl. Organomet. Chem. 16, 245–249 (2002).
- [39] Kuehnelt D., J. Lintschinger, W. Goessler: Arsenic compounds in terrestrial organisms. IV. Green plants and lichens from an old arsenic smelter site in Austria. Appl. Organometall. Chem. 14, 411–420 (2000).
- [40] Dercová K., J. Makovníková, G. Barančíková, J. Žuffa: Bioremediácia toxických kovov kontaminujúcich vody a pôdy. Chem. Listy 99, 682–693 (2005).
- [41] Iskandar I.K., M.B. Kirkham: Trace Elements in Soils. Lewis Publisher, CRC Press Boca Raton (2001).
- [42] Gonzaga M.I.S., J.A.G. Santos, L.Q. Ma: Arsenic phytoextraction and hyperaccumulation by fern species. Sci. agric. 63, 90–101 (2006).
- [43] Reddy C. N., W. H. Patrick: Effect of redox potential and pH on the uptake of cadmium and lead by rice plants. J. Environ. Qual. 6, 259–262 (1977).
- [44] Gavi F., T. Basta, W.R. Raun: Wheat grain cadmium as affected by long-term fertilization and soil acidity. J. Environ. Qual., 26, 265–271 (1997).
- [45] Alloway B.J.: Heavy Metals in Soils. Blackie and Son Ltd., Glasgow and London (1990).

Received: February 17, 2011; accepted: May 16, 2011.