

# 1 The potential of *Stenotrophomonas maltophilia* KB2 for phenol degradation 2 under exposure to heavy metal

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## 12 Abstract

13 The large diversity of chemical substances present in air, water, or soil makes it necessary to  
14 study their mutual impact on the effectiveness of microbiological decomposition of  
15 contaminants. This publication presents the results of the studies aimed at evaluating the effect  
16 of two biogenic heavy metals - zinc and copper - on the phenol biodegradation by the  
17 *Stenotrophomonas maltophilia* KB2 strain. The tests were carried out for concentrations of  
18 metals significantly exceeding the legally permitted wastewater values: for zinc up to  
19  $13.3 \text{ g}\cdot\text{m}^{-3}$ , and copper up to  $3.33 \text{ g}\cdot\text{m}^{-3}$ . In the tested metal concentration range, phenol  
20 biodegradation by the *S. maltophilia* KB2 strain was not significantly influenced by the  
21 introduced dose of zinc. While the presence of copper inhibited both biomass growth and  
22 substrate degradation. Kinetic data of metal and phenol mixtures were analyzed and very good  
23 correlations were obtained for the proposed equations. An equation consistent with the Han  
24 and Levenspiel model was proposed for the system *S. maltophilia* KB2-phenol-copper, while  
25 an equation consistent with the Kai model for the system *St. maltophilia* KB2-phenol-zinc.  
26 The simultaneous presence of Zn and Cu ions in the culture resulted in a stronger inhibition of  
27 phenol biodegradation.

28 **Keywords:** phenol, heavy metal, biodegradation, kinetic equations, synergism

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29

## 1. INTRODUCTION

30 The rapid development of industry and agriculture generate varieties of waste, increased water  
31 consumption and the amount of discharged wastewater affecting aquatic ecosystems. Owing to  
32 the use of various raw materials and complexity of processes, wastewater generally contains  
33 many inorganic and organic pollutants like polycyclic aromatic hydrocarbons, nitrogen,  
34 phosphorous and sulphur, heavy metals as well as other emerging and harmful substances  
35 (Gaurav 2021, Butarewicz 2019) Wastewater can be treated using various methods that are  
36 generally classified as physical, chemical or biological. These techniques have both advantages  
37 and disadvantages, however the primary goal is high efficiency of pollution removal, so they  
38 are constantly being improved (Ahmed et al., 2021, Bibi et al., 2023, Khan et al., 2023, Maziotis  
39 and Molinos-Senante, 2023). Biological techniques have been continuously developed for  
40 several decades, which has made them effective, environmentally friendly, simple, and  
41 inexpensive solutions in many cases (Khalidi-Idrissi et al., 2023; Mohd, 2020, Hussain et al.,  
42 2021, Tang et al., 2023). Microorganisms (bacteria, fungi, yeasts) have unique predispositions  
43 that enable them to use enzymes to decompose xenobiotic substances into CO<sub>2</sub>, H<sub>2</sub>O, SO<sub>2</sub>, NH<sub>3</sub>,  
44 or other small molecules, and to synthesize the organic matter they need (Panigrahy et al., 2022;  
45 Priyadarshini et al., 2021,2022). Good results are achieved by modifying the conventional  
46 biological systems conjoining with physical, chemical, and other biological systems in hybrid  
47 technologies (Ahmed et al., 2021, Saidulu et al., 2021).

48 Frequently detected pollutants are phenol and its derivatives occurring in post-production  
49 wastewater from the petrochemical, textile, pharmaceutical, plastic, and chemical industries  
50 (Gadipelly et al., 2014; Tutić et al., 2023). Although there are natural sources of phenol, it is a  
51 priority pollutant that shouldn't be discharged directly into the environment. To meet the  
52 demand for phenol, approximately 7 billion kilograms of this organic compound are produced  
53 annually (Chen and Sun, 2023). Due to the enormous scale of this basic organic raw material  
54 use, phenol occurs in the air, water, soil, and bottom sediments. It can be detected in most  
55 industries wastewaters in various concentrations, ranging from a minimal 1 mg·L<sup>-1</sup> to 7000  
56 mg·L<sup>-1</sup> (Mohd, 2020). Phenol is very toxic to neurons, the respiratory tract, the eyes, and the  
57 skin. Prolonged or repeated exposure to this organic substance may affect the condition and  
58 function of the kidneys and liver. Another mechanism for the toxicity of phenol may be the  
59 formation of phenoxyl radicals (Ataei et al., 2024). Undoubtedly, the effect of industrial  
60 wastewater treatment will be significantly influenced by the amount of phenol (Villegas et al.,  
61 2016), but also by the quantity and quality of accompanying substances, e.g. heavy metals.  
62 Some of the heavy metals (Fe, Cu, Co, Zn, Ni, Mn, Se, and Mo) are quintessential at low

63 concentrations for the normal physiological functions of living organisms. Specific amounts of  
64 them are required for the survival of organisms, while their high concentrations are toxic for  
65 humans, plants, and microorganisms. Other heavy metals, such as Ag, As, Cd, Pb, Hg, and  
66 Cr(VI) do not have a known biological function and are toxic in any quantity causing  
67 deleterious effects on the well-being and survival of living organisms (Gomathy and  
68 Sabarinathan, 2010; Abd Elnabi et al., 2023). Due to the ubiquity of metals in the environment,  
69 microorganisms have developed various ways of dealing with them, for example, active and  
70 passive uptake of metal ions, sequestration and immobilization of metals, chelation of metals  
71 outside the cell, solubilization by releasing organic acids (Sharma et al., 2023; Syed et al.,  
72 2023).

73 Copper is a micronutrient necessary for the proper functioning of humans and animals. The  
74 basic function is to participate in oxidation-reduction processes, where it occurs as a coenzyme  
75 and in the production of erythrocytes or the synthesis of hemoglobin. The sorption properties  
76 of soils towards copper are much higher than towards other trace cations. A constant increase  
77 in the content of this metal is observed especially in the orchard and horticultural soils, as well  
78 as near industrial areas, urban agglomerations, and near the road network (Abd Elnabi et al.,  
79 2023).

80 Zinc is an essential element fulfilling several basic functions in the metabolic processes of living  
81 organisms. However, large doses of zinc cause damage to many biochemical processes and are  
82 deposited in the kidneys, liver, and sex glands. Zinc is widely used mainly as a component of  
83 alloys, in pressure casting, printing, and shipbuilding industries. Additionally, it is used in the  
84 production of electric tools, as well as in the tool, lighting, and metal industries. Most of the  
85 zinc produced is used for galvanizing to protect iron and steel from rusting. The highest  
86 concentrations of this metal occur in soils affected by emissions from non-ferrous metal  
87 smelters, and zinc also enters agricultural soils with fertilizers (Mitra et al., 2022)

88 Both phenol and heavy metals have been included on the US Environmental Protection  
89 Agency's (US EPA) list of 126 priority environmental pollutants. According to the regulation  
90 of the Minister of Maritime Economy and Inland Navigation from 2019 (Dz. U. 2019 poz.  
91 1311) on the conditions to be met when discharging sewage into water or land, the highest  
92 permissible values of pollution indicators for industrial sewage are: for zinc  $2 \text{ mg} \cdot \text{dm}^{-3}$  (all  
93 types of wastewater), and copper  $0.1 \text{ mg} \cdot \text{dm}^{-3}$  (ceramics industry) and  $0.5 \text{ mg} \cdot \text{dm}^{-3}$  (other types  
94 of wastewater). Biochemical processes are inhibited at zinc concentrations above  $10 \text{ mg} \cdot \text{dm}^{-3}$ ,  
95 while nitrification processes are significantly inhibited at concentrations above  $2 \text{ mg} \cdot \text{dm}^{-3}$  of  
96 zinc.

97 Numerous bacteria, fungi and yeasts such as *Alcaligenes*, *Pseudomonas*, *Bacillus*,  
98 *Rhodococcus*, *Micrococcus*, *Cellulosimicrobium*, *Microbacterium*, *Flavobacterium*,  
99 *Methanospirillum*, *Aeromonas*, *Sphingobium*, *Aspergillus*, *Penicillium*, *Trichoderma*,  
100 *Streptomyces* and *Candida* have been isolated and characterized as detoxifying the  
101 contaminants (Zhao et al., 2021, Zhang et al., 2023). New microorganisms are still being  
102 sought, and their potential in the treatment of contaminated water is widely researched due to  
103 the benefits that biodegradation offers (Miglani et al., 2022; Abd Elnabi et al., 2022). However,  
104 not all microorganisms that can degrade xenobiotics under the settled optimal laboratory  
105 conditions can be successfully applied in the remediation of contaminated wastewater or soil.  
106 Many of them are sensitive to periodically changing environmental conditions (temperature,  
107 pH, oxygen concentration, salinity) or the presence of coexisting pollutants, such as heavy  
108 metals.

109 *Stenotrophomonas maltophilia* – an opportunistic pathogen widespread in the environment,  
110 which can cause, among others, respiratory and bloodstream infections also belongs to the  
111 group of bacteria with the ability to degrade organic compounds. An important feature of *S.*  
112 *maltophilia* is its ability to form biofilms on moist surfaces, which enhances its resistance to  
113 antimicrobials (Brooke, 2021; García et al., 2023). Its ability to produce enzymes,  
114 nanoparticles, and inhibitory molecules that are useful not only in environmental protection but  
115 also in food production and agriculture makes it of wide interest in biotechnology. The potential  
116 of *S. maltophilia* to degrade xenobiotics has been exploited for several years (Dias et al., 2022;  
117 Chen et al., 2014, 2016; Wu et al., 2021; Alvarado-Gutiérrez et al., 2020).

118 Although the microbial degradation of phenol is well documented, the impact of co-pollutants  
119 are still not exhaustively investigated and described. The problem is the diversity and quantity  
120 of organic and inorganic compounds occurring in the environment and the lack of information  
121 about their interactions, hence it is so important to investigate the mutual influence and their  
122 impact on the effectiveness of biodegradation (Goutam Mukherjee et al., 2022; Štefanac et al.,  
123 2021). Assessment of the influence of two biogenic heavy metals – zinc and copper – on the  
124 phenol biodegradation by the *Stenotrophomonas maltophilia* KB2 strain will be presented  
125 below. The results of this study could be useful e.g. in optimizing operational conditions of  
126 existing plants.

127

## 2. MATERIALS AND METHODS

128

**2.1. Chemicals and medium**

129 The bacteria lived in the mineral salts medium (MSM) containing: 3.78 g  $\text{Na}_2\text{HPO}_4 \times 12\text{H}_2\text{O}$ ;  
130 0.5 g  $\text{KH}_2\text{PO}_4$ ; 5 g  $\text{NH}_4\text{Cl}$ ; 0.2 g  $\text{MgSO}_4 \times 7\text{H}_2\text{O}$ ; 0.01 g yeast extract; deionized water 1000  
131 mL; enriched with 1 mL of TMS (Trace Mineral Solution). The phenol biodegradation in the  
132 presence of  $\text{Cu}^{2+}$  or  $\text{Zn}^{2+}$  ions was tested using a medium containing less phosphate salts to  
133 prevent precipitation. In preliminary tests, the optimal content of  $\text{Na}_2\text{HPO}_4 \times 12\text{H}_2\text{O}$  ( $1.26 \text{ g}\cdot\text{L}^{-1}$ )  
134 and  $\text{KH}_2\text{PO}_4$  ( $0.167 \text{ g}\cdot\text{L}^{-1}$ ) was determined. The metal solution was prepared by soluble  
135  $\text{ZnSO}_4 \times 7\text{H}_2\text{O}$  or  $\text{CuSO}_4 \times 5\text{H}_2\text{O}$  in deionized water.

136

**2.2. Chemical analysis**

137 The samples were collected from the bioreactor and centrifuged (15 000 rpm, 15 min, 4°C),  
138 filtered (2  $\mu\text{m}$  pore diameter), and diluted with deionized water. The changes of substrate  
139 concentration in the liquid culture were determined by chromatographic analyses (Waters  
140 HPLC equipped with Waters 1525 gradient pump and two-wave detector UV-VIS Waters  
141 M2487, a reverse phase column (Spherisorb ODS 2, 5  $\mu\text{m}$ , 150 $\times$ 4.6 mm), methanol and 1%  
142 acetic acid (40:60 v/v) with the flow rate of 1  $\text{mL}\cdot\text{min}^{-1}$ ); detection was carried out at  
143 wavelength 272 nm. Bacterial cell density was determined by measuring the absorbance at a  
144 wavelength  $\lambda = 550 \text{ nm}$  (spectrophotometer HACH DR3900). The metal ion concentration was  
145 checked using colorimetric analysis (cuvette tests, Hach) at the beginning and end of the  
146 experiment.

147

**2.3. Microorganisms**

148 The *Stenotrophomonas maltophilia* KB2 strain is stored under number E-113197 in the VTT  
149 Collection in Finland. This strain was isolated from activated sludge of the wastewater  
150 treatment plant in Bytom – Miechowice, Poland, as described earlier (Guzik et al., 2009). It is  
151 known from previous studies that the KB2 strain has the ability to utilize different aromatic  
152 substrates as sole carbon and energy source, e.g. phenol, catechol, benzoic acid, protocatechuic  
153 acid, 4-hydroxybenzoic acid, and vanillic acid (Guzik et al., 2009). Guzik et al., 2012  
154 demonstrated that strain KB2 was able to utilize  $1100 \text{ g}\cdot\text{m}^{-3}$  phenol during 24 h of incubation.  
155 The abilities of *S. maltophilia* KB2 to degrade phenol effectively under suboptimal temperature,  
156 pH, and salinity were confirmed in Nowak et al., 2022. Tests on the sensitivity of catechol 2,3-  
157 dioxygenase isolated from *Stenotrophomonas maltophilia* KB2 cells demonstrated that the

158 activity of this enzyme increased in the presence of  $Zn^{2+}$  ions by 18% but decreased to 0.5% in  
159 the presence of copper ions. Phenolic monooxygenase lost its activity after adding copper  
160 sulphate or iron chloride (Guzik et al., 2012).

161 The preparation of microorganisms for the kinetics studies was described in the publication  
162 Gąszczak et al., 2021.

#### 163 ***2.4. Biodegradation experiments***

164 Biodegradation tests were carried out in the stirred batch bioreactor (Biostat B fermenter,  
165 Sartorius, USA) equipped with temperature, pH, and  $O_2$  sensors.

166 The concentration of dissolved oxygen (DO) in the suspension was kept at  $5 \text{ mg} \cdot \text{dm}^{-3}$  (external  
167 compressor). The other environmental conditions were:  $30^\circ\text{C}$ ,  $\text{pH}=7$ , and stirrer rotations at 300  
168 rpm. Each cultivation was started with the  $\text{OD}=0.2$  which was equaling  $61.3 \text{ g}_{\text{dew}} \cdot \text{m}^{-3}$ . The  
169 culture was sampled at regular time intervals, only the metal concentration was checked at the  
170 beginning and end of the test.

#### 171 ***2.5. Modelling kinetics of organic compound removal in the presence of metal***

172 The effect of heavy metal ions presence on the bacterial growth rate or organic compound  
173 degradation rate is widely discussed in the literature. However, despite the importance of such  
174 research, the presented mathematical descriptions of these phenomena are insufficient. Several  
175 mathematical models that are developments of the Monod equation, have been found in the  
176 literature.

177 The Han and Levenspiel model considers the existence of a critical inhibitor concentration,  
178 above which the growth rate slows down or the reaction stops completely. (Gąszczak et al.,  
179 2021). The modified version of this model:

$$180 \quad \mu = \mu_{max} \left(1 - \frac{M}{M_{crit}}\right)^n \quad (1)$$

181 where  $M$  is the metal concentration, and  $M_{crit}$  is the critical metal concentration was applied to  
182 describe the organic compound biodegradation in an environment containing heavy metals  
183 (Gopinath et al., 2011; Shukor et al., 2018; Manogaran et al., 2019).

184 To describe the growth of *Rhodococcus* sp. n diesel fuel in the presence of zinc Kai et al. (Kai  
185 et al., 2020) used the equation:

$$186 \quad \mu = \mu_{max} \cdot e^{-k \cdot M} \quad (2)$$

187 Equation 3 accounts for both the substrate inhibition and heavy metal ion inhibition:

$$\mu = \frac{\mu_{max} \cdot S}{K_S + S + \frac{S^2}{K_{IS}}} \cdot \frac{K_{IM}}{K_{IM} + M^n} \quad (3)$$

where  $K_{IS}$  represents the substrate inhibition constant,  $K_{IM}$  represents the heavy metal inhibition constant, and  $n$  is an equation parameter. Constants  $K_{IM}$  and  $n$  can be determined graphically

$$\left( \log \left[ \frac{\mu_{max} \cdot S}{\mu \left( K_S + S + \frac{S^2}{K_{IS}} \right)} - 1 \right] \right) = f(\log M) \quad (\text{Nakamura and Sawada, 2000}).$$

Amor et al. used a simplified form of the Andrews equation (believing that  $K_S$  was irrelevant) to model the effect of cadmium, nickel, and zinc on rates of alkyl benzene biodegradation (Amor et al., 2001).

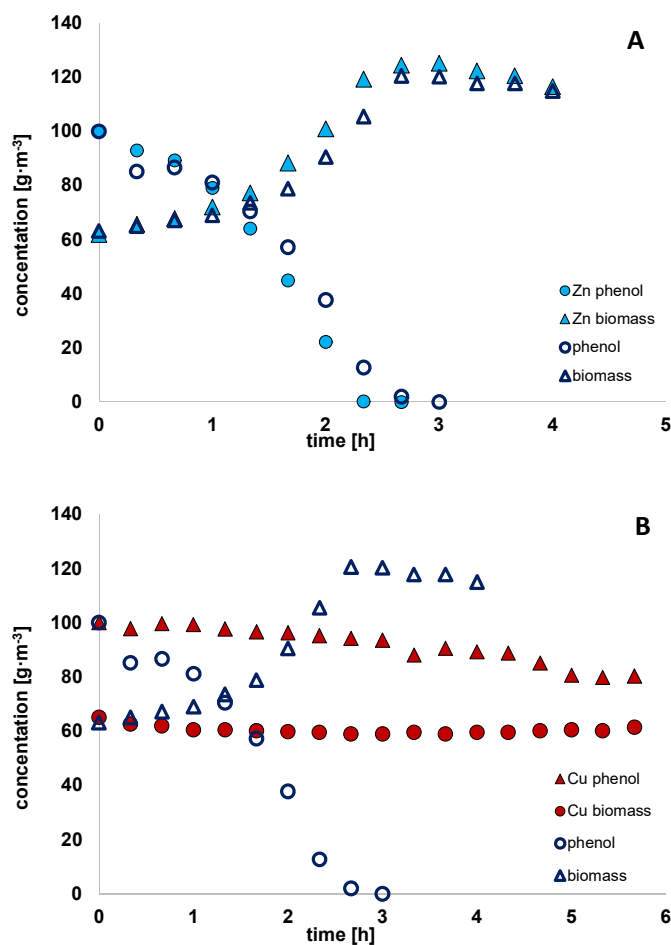
$$\frac{1}{\mu} = \frac{1}{\mu_{max}} + \frac{M}{K_{IM} \cdot \mu_{max}} \quad (4)$$

### 3. RESULTS

The large diversity of chemical substances released into the natural environment makes it necessary to investigate the interaction of organic and inorganic compounds and their influence on the effectiveness microbial decomposition of contaminants.

#### 3.1. Phenol biodegradation

The frequently detected contaminants include phenol, its derivatives, and heavy metals. The kinetics of the phenol biodegradation by *Stenotrophomonas maltophilia* KB2 were studied previously and the results were described in the paper Gąszczak et al., 2021. The following values of the Andrews equation parameters were reported:  $\mu_m = 1.584 \text{ h}^{-1}$ ,  $K_S = 185.4 \text{ g} \cdot \text{m}^{-3}$ ,  $K_{IS} = 106.1 \text{ g} \cdot \text{m}^{-3}$ .



206

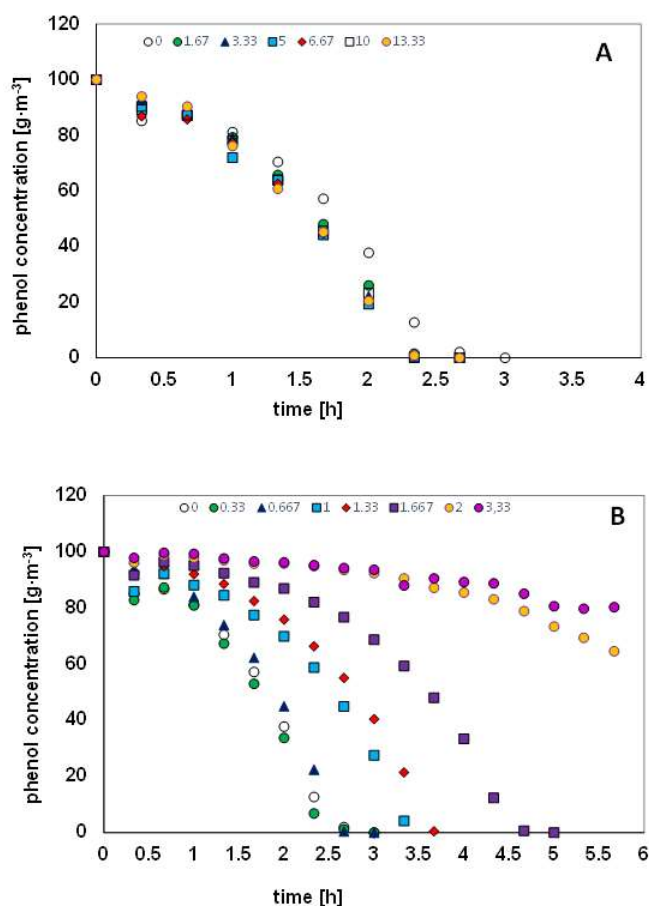
207 Fig.1. Example cultures with metal at  $3.33 \text{ g}\cdot\text{m}^{-3}$  (full tracers) and without (empty tracers);  
 208 circles – phenol concentration, triangles – biomass concentration; A) zinc, B) copper.

209

210 The tolerance of *S. maltophilia* KB2 strain to zinc and copper was assessed by evaluating  
 211 changes in phenol and biomass concentrations in culture media amended with various  
 212 concentrations of heavy metal ions. Two series of tests were carried out for the zinc  
 213 concentrations ranging from  $1.67$  to  $13.33 \text{ g}\cdot\text{m}^{-3}$  and for copper concentrations ranging from  
 214  $0.33$  to  $3.33 \text{ g}\cdot\text{m}^{-3}$ , at the initial phenol concentration of  $100 \text{ g}\cdot\text{m}^{-3}$ . As shown in a previous  
 215 publication (Gąszczak et al., 2021)  $100 \text{ g}\cdot\text{m}^{-3}$  is the phenol concentration at which the specific  
 216 growth rate is close to the maximum value. Figure 1 shows the courses of exemplary cultures  
 217 in the presence of zinc (A) or copper (B) at a concentration of  $3.33 \text{ g}\cdot\text{m}^{-3}$ .

218 Even though both metals belong to the group of biogenic metals, their presence in the culture  
 219 medium caused different effects. The presence of zinc, in the tested concentration range, had a  
 220 slight effect on the course of phenol degradation, which was accelerated by several minutes  
 221 (Figure 2A). Both the efficiency of phenol degradation and biomass growth decreased as the  
 222 copper concentration increased. This relationship is illustrated in Figure 2B.





223

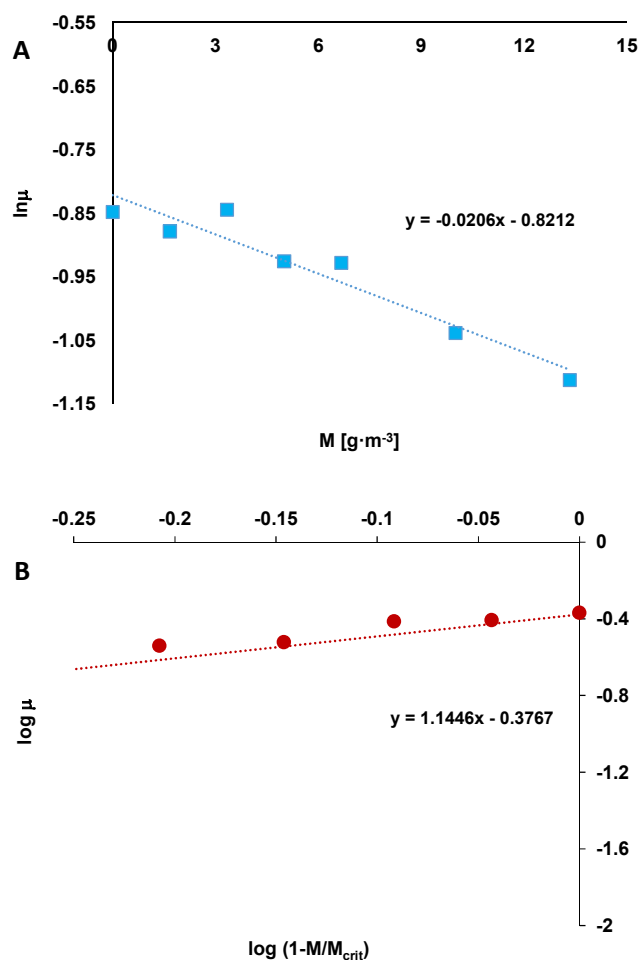
224 Fig. 2. The effect of the metal concentrations [g·m<sup>-3</sup>] on phenol utilization; A zinc and B  
 225 copper.

226 Copper turned out to be a strong inhibitor of phenol biodegradation by the KB2 strain. Small  
 227 concentrations significantly slow down this process and even at a concentration of 1 g·m<sup>-3</sup>, the  
 228 phenol degradation time was significantly prolonged. The highest copper concentration tested  
 229 was 3.33 g·m<sup>-3</sup>. This dose of copper resulted in a negligible, in relation to the size of the  
 230 inoculum, biomass growth, which was practically stopped. It can be assumed that the  
 231 accumulation of Cu<sup>2+</sup> ions in cells inhibited the activity of enzymes: catechol 2,3-dioxygenase  
 232 and protocatechuate 3,4-dioxygenase (Guzik et al., 2009, Silva et al., 2012).

### 233 *3.2. Modeling the Specific Growth Rate as a Function of Different Metal* 234 *Concentrations*

235 To study the kinetics of heavy metal inhibition, the phenol biodegradation experiments were  
 236 carried out for various metal concentrations in a batch bioreactor. The specific growth rate was  
 237 determined for each culture. The established experimental database was used to estimate the  
 238 equation parameters for selected mathematical models (Equations 1-4). Methods for

239 determining the parameters of the above-mentioned equations are described in the publication  
 240 (Gąszczak et al., 2021).



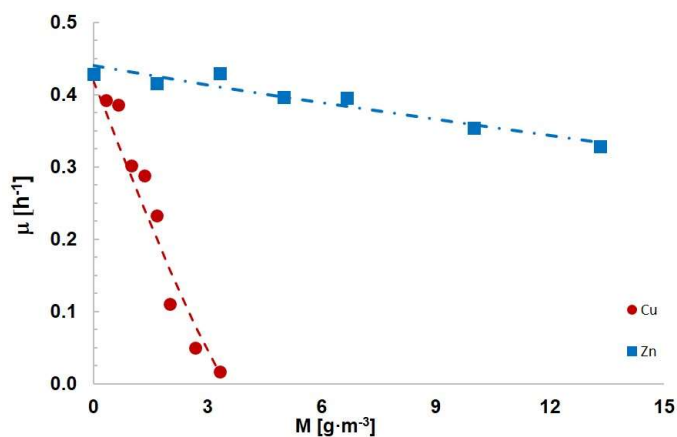
241  
 242 Fig. 3. The method of determining the parameters of the Kai model (A) and Han-Levenspiel  
 243 model (B).  
 244 The best fit of the model and experimental data was obtained: for copper – with the Han-  
 245 Levenspiel model, and for zinc – with the Kai model. Figure 3 illustrates the determination of  
 246 the parameters of these models.

247 Kai model for Zn: 
$$\mu = 0.44 \cdot e^{-0.0206 \cdot M} \quad (5)$$

248 Han-Levenspiel model for Cu: 
$$\mu = 0.42 \left(1 - \frac{M}{3.5}\right)^{1.1446} \quad (6)$$

249 Both Kai and Han-Levenspiel models showed good fitting:  $R^2=0.924$  and  $R^2=0.948$ ,  
 250 respectively.

251 Figure 4 shows the inhibitory effect of increasing metal concentrations on the specific growth  
252 rate of *S. maltophilia* KB2 strain, the experimental points, and fitted the Han–Levenspiel and  
253 Kai models.



254

255 Fig. 4. Inhibitory effect of increasing concentrations of copper or zinc to the specific growth  
256 rate of *S. maltophilia* KB2 strain; phenol concentration 100 g·m<sup>-3</sup> (●) Cu, (■) Zn.

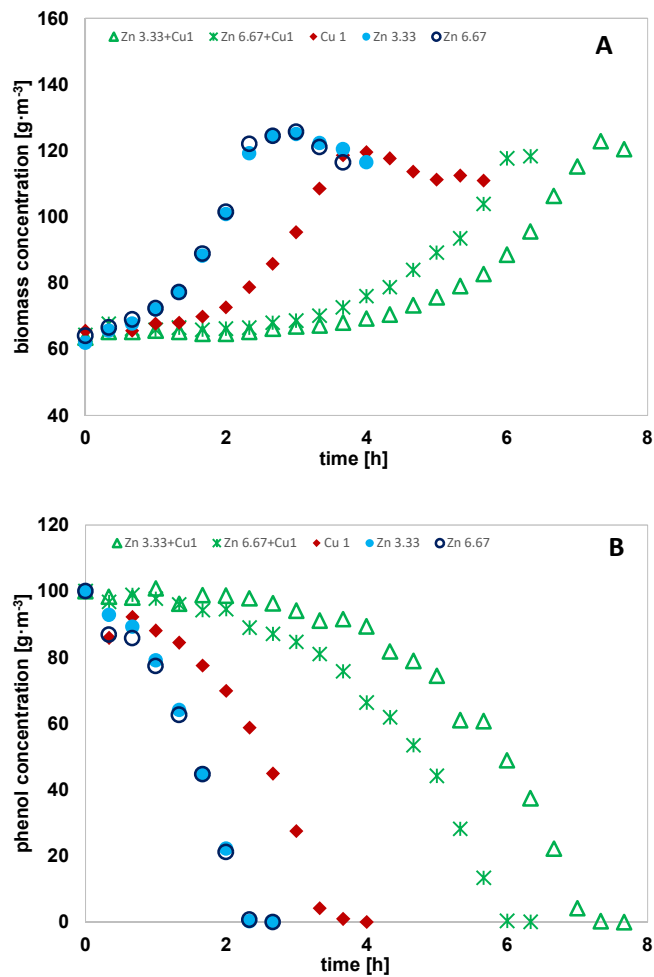
257

### 258 3.3. The Effect of Mixed Heavy Metals on the Phenol Degradation

259 Synergism refers to a stronger toxic response from exposure to two or more chemicals than  
260 would be expected based on the sum of the effects of the individual chemicals. Synergistic  
261 properties should be taken into consideration when assessing the potential hazards of chemicals.

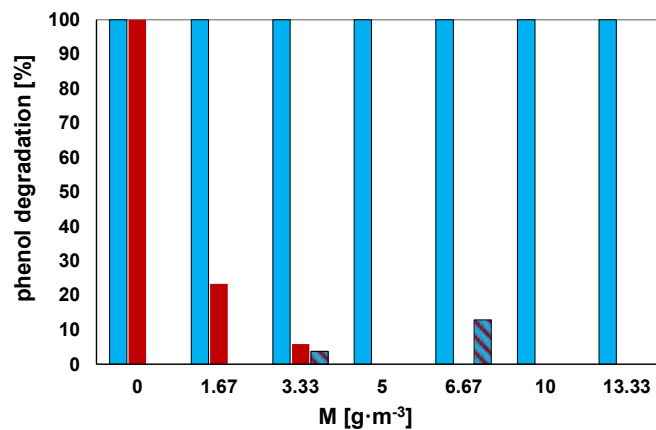
262 A similar phenomenon is potentiation but in this case, one of the chemicals has no apparent  
263 ability to produce the toxic response on its own. In contrast, antagonism occurs when two or  
264 more substances in the mixture cause a weaker effect than when acting individually.

265 Since heavy metals are more likely to be present in a common environment as mixed heavy  
266 metals than as single ones, an attempt was made to analyse the effect of mixed zinc and copper  
267 on phenol degradation. Figure 5 shows the biomass growth and phenol degradation depending  
268 on the presence of single or mixed heavy metals. In the mixture, the copper concentration was  
269 1 g·m<sup>-3</sup>, while the zinc content was 3.33 or 6.67 g·m<sup>-3</sup>.



270  
 271  
 272

Fig. 5. The effect of mixed heavy metals on the phenol degradation



273  
 274  
 275

Fig. 6. The effect of metal concentration on phenol degradation at  $100 \text{ g}\cdot\text{m}^{-3}$ ; Zn (blue bars), Cu (red bars), and Zn+Cu (blue/red pattern bars).

276 During the experiments, a much stronger toxic effect was observed after the introduction of a  
 277 mixture of Zn and Cu than in cultures containing only one metal (Fig.5). The graph (Figure 6)

278 shows a comparison of the impact of the presence of zinc and copper ions on the phenol  
279 biodegradation. A dose of  $100 \text{ g}\cdot\text{m}^{-3}$  was utilized in 100% by the *S.maltophilia* KB2 strain  
280 within 140 minutes. In the tested range of zinc concentrations, the degradation time of this dose  
281 of phenol did not change. As already mentioned, copper turned out to be a strong inhibitor of  
282 this process. Only 58.8% of the substrate was utilized at the same period (140 minutes), at a  
283 copper concentration of  $1 \text{ g}\cdot\text{m}^{-3}$ , while at the highest tested concentration of copper ions  
284 ( $C_{\text{Cu}}=3.33 \text{ g}\cdot\text{m}^{-3}$ ) only 5.8%. When, except copper at a concentration of  $1 \text{ g}\cdot\text{m}^{-3}$ , zinc was also  
285 present in the culture medium, the obtained percentage of utilized phenol, depending on zinc  
286 concentration, was: 3,7% ( $C_{\text{Zn}}=3.33 \text{ g}\cdot\text{m}^{-3}$ ) and 12,9 ( $C_{\text{Zn}}=6.67 \text{ g}\cdot\text{m}^{-3}$ ). Noteworthy is the fact  
287 that a higher concentration of zinc resulted in a greater amount of utilized phenol at the same  
288 time. Is this phenomenon synergism or potentiation? The key seems to be the interpretation of  
289 the effects induced by the presence of zinc in culture. Considering Figure 2A, it can be  
290 concluded that zinc is not toxic to *S. maltophilia* KB2, so it would be a potentiation. However,  
291 in Figure 4 you can see a decrease in the value of the specific growth rate with increasing zinc  
292 concentration. Therefore, it seems justified to call the effect caused by the simultaneous  
293 introduction of copper and zinc ions synergism. To better understand this phenomenon, more  
294 research needs to be conducted.

295 Most works on environmental pollution by heavy metals describe their harmfulness and  
296 removal methods. Much fewer publications have been devoted to the interaction of pollutants,  
297 e.g. xenobiotics + heavy metals, and the problems occurring during their removal. Nakamura  
298 and Sawada studied phenol biodegradation by *Acinetobacter calcoaceticus* AH strain in a  
299 solution containing heavy metals such as zinc or copper ions. The model they proposed could  
300 satisfactorily express the inhibitory effects of substrate and heavy metals and it showed fair  
301 agreement with the experimental data. Similar research was carried out by Zhang et al. (Zhang  
302 et al., 2022), who isolated a new strain of *Bacillus cereus* ZBW3. The addition of  $\text{Zn}^{2+}$  slowed  
303 down the rate of phenol degradation, however, the complete degradation occurred within 72  
304 hours. Conversely, the presence of  $\text{Cu}^{2+}$  ions resulted in the complete inhibition of phenol  
305 degradation, as their accumulation in cells hindered the activity of catechol dioxygenase (Zhang  
306 et al., 2022). The co-culture consisting of *Arthrobacter* sp. strain AQ5-15 and *Arthrobacter* sp.  
307 strain AQ5-06 exhibited phenol degradation capabilities up to 1.7 g/L within a week. The  
308 examination of resistance to heavy metals found that the disintegration of phenol by this co-  
309 culture was completed in the presence of As, Al, Co, Cr Cu, Ni, Pb, and Zn at 1.0 ppm  
310 concentrations. However, the activity of *Arthrobacter* strains was inhibited by Cd, Ag, and Hg  
311 (Subramaniam et al., 2021).

312 A literature review of the effects of pollutant co-interactions showed that they depend on the  
313 nature of the pollutants and their concentrations or proportions in the mixture (Nlemolisa et al.,  
314 2020). The research by Batkhuyag et al. (2021) investigated the additive inhibitory effects of  
315 heavy metals on the utilization of phenol by various microorganisms, namely *Alicyclophilus*  
316 *denitrificans* K601, *Alicyclophilus* sp. R-2461, uncultured *Alicyclophilus* sp., and *Acidovorax*  
317 *aerodenitrificans*. The inhibitory effects of binary heavy metal mixtures on phenol-using  
318 microorganisms were found to follow a particular order based on synergistic interactions. This  
319 order was as follows: (Cd + Pb)  $\approx$  (Cd + Cu) > (Zn + Pb) > (Zn + Cd) > (Pb + Cu) > (Zn + Cu).

320

#### 4. CONCLUSIONS

321 What is the impact of heavy metals presence on the degradation of phenol by the  
322 *Stenotrophomonas maltophilia* KB2 strain? Due to such different courses of the processes  
323 depending on the introduced element, it is not possible to give a concise answer to this question.  
324 The slight increase in copper concentration resulted in a significant decrease in phenol  
325 degradation efficiency, whereas higher zinc concentrations had no practical effect on phenol  
326 biodegradation. The most appropriate equation to describe the system *S. maltophilia* KB2-  
327 phenol-copper utilises the Han and Levenspiel model. Likewise, for the system *S. maltophilia*  
328 KB2-phenol-zinc, an equation consistent with the Kai model was found to be the most suitable.  
329 The simultaneous presence of Zn and Cu ions in the culture resulted in a stronger inhibition of  
330 phenol biodegradation. At present, there is no generally accepted kinetic equation to describe  
331 the multiple effects of the introduction of co-pollutants. However, there is an urgent need to  
332 develop a new model for microbial growth and pollutant degradation that takes into account  
333 multiple substrates and inhibitors in an actual industrial setting. Understanding the  
334 biodegradation requirements and kinetics of microbial growth is imperative for designing and  
335 scaling up efficient bioreactor systems and optimizing technological processes to meet high  
336 quality standards.

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#### REFERENCES

- 339 1. Abd Elnabi M.K., Elkaliny N.E., Elyazied M.M., Azab S.H., Elkhalfifa S.A., Elmasry S.,  
340 Mouhamed M.S., Shalamesh E.M., Alhorienny N.A., Abd Elaty A.E.; et al.,2023.Toxicity  
341 of Heavy Metals and Recent Advances in Their Removal: A Review. *Toxics*, 11, 580. DOI:  
342 10.3390/toxics11070580

- 343 2. Ahmed S.F., Mofijur M., Nuzhat S., Chowdhury A.T., Rafa N., Alhaz Uddin Md., Inayat  
344 A., Mahlia T.M.I., Ong H., Chia W.Y., Show P., 2021. Recent developments in physical,  
345 biological, chemical, and hybrid treatment techniques for removing emerging contaminants  
346 from wastewater. *Journal of Hazardous Materials*, 416, 125912,  
347 DOI:10.1016/j.jhazmat.2021.125912.
- 348 3. Alvarado-Gutiérrez M.L., Ruiz-Ordaz N., Galíndez-Mayer J., Curiel-Quesada E., Santoyo-  
349 Tepole F., 2020. Degradation kinetics of carbendazim by *Klebsiella oxytoca*,  
350 *Flavobacterium johnsoniae*, and *Stenotrophomonas maltophilia* strains. *Environmental*  
351 *Science and Pollution Research*, 27, 28518–28526. DOI:10.1007/s11356-019-07069-8.
- 352 4. Amor L., Kennes C., Veiga M.C., 2001. Kinetics of inhibition in the biodegradation of  
353 monoaromatic hydrocarbons in the presence of heavy metals. *Bioresource Technology*, 78,  
354 181–185. DOI: 10.1016/S0960-8524(00)00182-6.
- 355 5. Ataei M, Maghsoudi A.S., Hassani Sh., 2024. *Phenol*, In: Editor(s): Philip Wexler,  
356 *Encyclopedia of Toxicology*, Fourth Edition, Academic Press, Pages 521-526, ISBN  
357 9780323854344, DOI: 10.1016/B978-0-12-824315-2.00168-8.
- 358 6. Batkhuyag N., Matyakubov B., Mang N.Z.L., Lee T., 2021. Additive inhibitory effects of  
359 heavy metals on phenol-utilizing microorganisms. *Environmental Engineering Research*,  
360 27, 210342–0. DOI: 10.4491/eer.2021.342.
- 361 7. Bibi A., Bibi Sh., Abu-Dieyeh M., Mohammad A. Al-Ghouti M., 2023. Towards  
362 sustainable physiochemical and biological techniques for the remediation of phenol from  
363 wastewater: A review on current applications and removal mechanisms. *Journal of Cleaner*  
364 *Production*, 417, 137810, DOI: 10.1016/j.jclepro.2023.137810.
- 365 8. Brooke, J.S., 2021. Advances in the microbiology of *Stenotrophomonas maltophilia*. *Clin.*  
366 *Microbiol. Rev.*, 34, e00030-19. DOI: 10.1128/CMR.00030-19.
- 367 9. Butarewicz A., Rosochacki S. J., Wrzaszcz E., 2019. Toxicity of sewage from industrial  
368 wastewater treatment plants. *Journal of Ecological Engineering*, 20(2), 191-199. DOI:  
369 10.12911/22998993/99060.
- 370 10. Chen S., Sun L., 2023. Screening of efficient phenol-degrading bacteria and analysis of  
371 their degradation characteristics. *Sustainability*, 15, 6788. DOI: 10.3390/su15086788
- 372 11. Chen S., Yin H., Tang S., Peng H., Liu Z., Dang Z., 2016. Metabolic biotransformation of  
373 copper–benzo[a]pyrene combined pollutant on the cellular interface of *Stenotrophomonas*  
374 *maltophilia*. *Bioresource Technology*, 204, 26–31. DOI: 10.1016/j.biortech.2015.12.068
- 375 12. Chen S., Yin H., Ye J., Peng H., Liu Z., Dang Z., Chang J., 2014. Influence of co-existed  
376 benzo[a]pyrene and copper on the cellular characteristics of *Stenotrophomonas maltophilia*

- 377 during biodegradation and transformation. *Bioresource Technology*, 158, 181–187. DOI:  
378 10.1016/j.biortech.2014.02.020
- 379 13. Dias P.R.P., Paiva T.O., de Oliveira A.M., de Magalhães J.C., 2022. Biodegradation of  
380 phenol by *Pseudomonas aeruginosa*, *Acinetobacter* sp. And *Stenotrophomonas*  
381 *maltophilia* isolated of the sludge activated of a steel industry, *International Journal of*  
382 *Development Research*, 12, (04), 55571-55574.
- 383 14. Dz. U. 2019 poz. 1311. Rozporządzenie Ministra Gospodarki Morskiej i Żegluggi  
384 Śródlądowej z dnia 12 lipca 2019 r. w sprawie substancji szczególnie szkodliwych dla  
385 środowiska wodnego oraz warunków, jakie należy spełnić przy wprowadzaniu do wód lub  
386 do ziemi ścieków, a także przy odprowadzaniu wód opadowych lub roztopowych do wód  
387 lub do urządzeń wodnych. Available at:  
388 <https://isap.sejm.gov.pl/isap.nsf/download.xsp/WDU20190001311/O/D20191311.pdf>
- 389 15. Gadipelly C., Pérez-González A., Yadav G.D., Ortiz I., Ibáñez R., Rathod V.K., Marathe  
390 K.V., 2014. Pharmaceutical Industry Wastewater: Review of the Technologies for Water  
391 Treatment and Reuse. *Ind. Eng. Chem. Res.* 53, 11571–11592. DOI: 10.1021/ie501210j
- 392 16. García G., Girón J.A., Yañez J.A., Cedillo M.L., 2022. *Stenotrophomonas maltophilia* and  
393 Its Ability to Form Biofilms. *Microbiology Research* 14, 1–20. DOI:  
394 10.3390/microbiolres14010001
- 395 17. Gomathy M., Sabarinathan K.G., 2010. Microbial mechanisms of heavy metal tolerance-  
396 a review. *Agricultural Reviews* 31 (2), 133-138
- 397 18. Gopinath K.P., Kathiravan M.N., Srinivasan R., Sankaranarayanan S., 2011. Evaluation  
398 and elimination of inhibitory effects of salts and heavy metal ions on biodegradation of  
399 Congo red by *Pseudomonas* sp. Mutant. *Bioresource Technology*, 102, 3687–3693. DOI:  
400 10.1016/j.biortech.2010.11.072
- 401 19. Goutam Mukherjee A., Ramesh Wanjari U., Eladl M.A., El-Sherbiny M., Elsherbini  
402 D.M.A., Sukumar A., Kannampuzha S., Ravichandran M., Renu K., Vellingiri B.,  
403 Kandasamy S., Valsala Gopalakrishnan A., 2022. Mixed Contaminants: Occurrence,  
404 Interactions, Toxicity, Detection, and Remediation. *Molecules*, 27, 2577. DOI:  
405 10.3390/molecules27082577
- 406 20. Guzik U., Greń I., Wojcieszńska D., Łabużek S., 2009. Isolation and characterization of  
407 a novel strain of *Stenotrophomonas maltophilia* possessing various dioxygenases for  
408 monocyclic hydrocarbon degradation. *Braz J Microbiol.*, 40, 285-291.
- 409 21. Guzik, U., Hupert-Kocurek, K., Sałek, K., Wojcieszńska, D., 2012. Influence of metal  
410 ions on bioremediation activity of protocatechuate 3,4-dioxygenase from



- 411 *Stenotrophomonas maltophilia* KB2. *World J. Microbiol. Biotechnol.*, 29, 267–273. DOI:  
412 10.1007/s11274-012-1178-z
- 413 22. Guzik U.; Wojcieszynska D.; Greń I.; Hupert-Kocurek K., 2010. Activity of Catechol  
414 Dioxygenases in the Presence of Some Heavy Metal Ions: Bioremediation of an  
415 Environment Polluted with Aromatic Compounds. *Ochrona Środowiska* , 32, 9–13
- 416 23. Hussain A., Kumari R., Sachan S.G., Sachan A., 2021. Biological wastewater treatment  
417 technology: advancement and drawbacks. *Microb. Ecol. Wastewater Treat. Plants*, 175-  
418 192, DOI: 10.1016/B978-0-12-822503-5.00002-3
- 419 24. Kai E.X.; Johari W.L.W.; Habib S.; Adeela N.; Ahmad S.A.; Shukor M.Y., 2020. The  
420 growth of *Rhodococcus* Sp. on diesel fuel under the effect of heavy metals and different  
421 concentrations of zinc. *Adv. Polar Sci.*, 31, 132–136
- 422 25. Khalidi-Idrissi A, Madinzi A, Anouzla A, Pala A, Mouhir L, Kadmi Y, Souabi S., 2023.  
423 Recent advances in the biological treatment of wastewater rich in emerging pollutants  
424 produced by pharmaceutical industrial discharges. *Int. J. Environ. Sci. Technol.* (Tehran).  
425 16,1-22. DOI: 10.1007/s13762-023-04867-z.
- 426 26. Khan N., López-Maldonado E., Majumder A., Singh S., Varshney R., J.R. López, P.F.  
427 Méndez, Ramamurthy P., Khan M., Khan A., Mubarak N., Amhad W., Shamshuddin S.,  
428 Aljundi I., 2023, A state-of-art-review on emerging contaminants: Environmental  
429 chemistry, health effect, and modern treatment methods. *Chemosphere*, 344, 140264, DOI:  
430 10.1016/j.chemosphere.2023.140264.
- 431 27. Manogaran M., Othman A.R., Shukor M.Y., Halmi M.I.E., 2019. Modelling the Effect of  
432 Heavy Metal on the Growth Rate of an SDS-degrading *Pseudomonas sp.* Strain DRY15  
433 from Antarctic soil. *Bioremed. Sci. Technol. Res.*, 7, 41–45. doi.org/10.54987/bstr.v7i1.463
- 434 28. Maziotis A. and Molinos-Senante M., 2023. A comprehensive eco-efficiency analysis of  
435 wastewater treatment plants: estimation of optimal operational costs and greenhouse gas  
436 emissions. *Water Research*, 243,120354, DOI: 10.1016/j.watres.2023.120354.
- 437 29. Mitra S., Chakraborty A., Tareq A., Emran T., Nainu F., Khusro A., Idris A., Khandaker M.,  
438 Osman H., Alhumaydhi F., Simal-Gandara J., 2022. Impact of heavy metals on the  
439 environment and human health: Novel therapeutic insights to counter the toxicity. *Journal*  
440 *of King Saud University - Science*, 34, 2022, 101865,  
441 doi.org/10.1016/j.jksus.2022.101865.
- 442 30. Miglani R., Parveen N., Kumar A., Ansari, Mohd A., Khanna S., Rawat G., Panda A.K.,  
443 Bisht S.S., Upadhyay J., Ansari M.N., 2022. Degradation of Xenobiotic Pollutants: An

- 444 Environmentally Sustainable Approach. *Metabolites* 12, 818. DOI:  
445 10.3390/metabo12090818
- 446 31. Mohd A., 2020. Presence of phenol in wastewater effluent and its removal: an overview.  
447 *International Journal of Environmental Analytical Chemistry*, 102, 1362–1384. DOI:  
448 10.1080/03067319.2020.1738412
- 449 32. Nakamura Y., Sawada T., 2000. Biodegradation of phenol in the presence of heavy metals.  
450 *J. Chem. Technol. Biotechnol.*, 75, 137–142. DOI: 10.1002/(SICI)1097-  
451 4660(200002)75:2<137::AID-JCTB194>3.0.CO;2-0
- 452 33. Nlemolisa O.R., Nwanyanwu C.E., Akujobi C.O., Ihenetu F.C., Nwokorie R.C., Obasi  
453 C.C., Kemka U.N., Uzoho K.H., Nwoke M.C., 2020. Toxicity of Binary Mixtures of  
454 Phenol, Zinc, and Cadmium to Yeast Strains Isolated from Hydrocarbon Impacted Soil.  
455 *OALib* 07, e6201,1–15. DOI: 10.4236/oalib.1106201
- 456 34. Nowak A., Wasilkowski D., Mroziak A., 2022. Implications of Bacterial Adaptation to  
457 Phenol Degradation under Suboptimal Culture Conditions Involving *Stenotrophomonas*  
458 *maltophilia* KB2 and *Pseudomonas moorei* KB4. *Water*, 14, 2845. DOI:  
459 10.3390/w14182845
- 460 35. Panigrahy N., Priyadarshini A., Sahoo M.M., Verma A.K., Daverey A., Sahoo N.K., 2022.  
461 A comprehensive review on eco-toxicity and biodegradation of phenolics: Recent progress  
462 and future outlook. *Environmental Technology & Innovation*, 27, 102423. DOI:  
463 10.1016/j.eti.2022.102423
- 464 36. Priyadarshini A., Mishra S., Sahoo M.M., Rout P.R., Sahoo N.K., 2022. Effect of nutrient  
465 and culture conditions on enhanced biodegradation of phenolic pollutants: A review on  
466 recent development and future prospective. *Environmental Quality Mgmt*, 32, 161–176.  
467 DOI: 10.1002/tqem.21934
- 468 37. Priyadarshini A., Sahoo M.M., Raut P.R., Mahant, B., Sahoo N.K., 2021. Kinetic modelling  
469 and process engineering of phenolics microbial and enzymatic biodegradation: A current  
470 outlook and challenges. *Journal of Water Process Engineering*, 44, 102421. DOI:  
471 10.1016/j.jwpe.2021.102421
- 472 38. Saidulu D., Gupta B., Gupta A., Ghosal P., 2021. A review on occurrences, eco-toxic  
473 effects, and remediation of emerging contaminants from wastewater: Special emphasis on  
474 biological treatment based hybrid systems. *Journal of Environmental Chemical*  
475 *Engineering*, 9, 4,2021,105282, DOI: 10.1016/j.jece.2021.105282.
- 476 39. Sharma M., Agarwal S., Agarwal Malik R., Kumar G., Pal D.B., Mandal M., Sarkar A.,  
477 Bantun F., Haque S., Singh P., Srivastava N., Gupta V.K., 2023. Recent advances in

- 478 microbial engineering approaches for wastewater treatment: a review. *Bioengineered*, 14,  
479 2184518. DOI: 10.1080/21655979.2023.2184518
- 480 40. Shukor M.Y., Gusmanizar N., Rusnam, 2018. Modelling the Effect of Heavy Metals on the  
481 Growth Rate of *Enterobacter* sp. Strain Neni-13 on SDS. *J Environ Microbiol Toxicol*, 6,  
482 24–27. DOI: 10.54987/jemat.v6i1.403
- 483 41. Silva A. S., Camargo F. A. D. O., Andrezza R., Jacques R. J. S., Baldoni D. B., & Bento  
484 F. M., 2012. Enzymatic activity of catechol 1, 2-dioxygenase and catechol 2, 3-  
485 dioxygenase produced by *Gordonia polyisoprenivorans*. *Quimica Nova*, 35, 1587-1592.
- 486 42. Subramaniam K., Ahmad S.A., Convey P., Shaharuddin N.A., Khalil K.A., Tengku-Mazuki  
487 T.A., Gomez-Fuentes C., Zulkharnain A., 2021. Statistical Assessment of Phenol  
488 Biodegradation by a Metal-Tolerant Binary Consortium of Indigenous Antarctic Bacteria.  
489 *Diversity* 13, 643. DOI: 10.3390/d13120643
- 490 43. Syed Z., Sogani M., Rajvanshi J., Sonu K., 2023. Microbial Biofilms for Environmental  
491 Bioremediation of Heavy Metals: a Review. *Appl Biochem Biotechnol*, 195, 5693–5711.  
492 DOI: 10.1007/s12010-022-04276-x
- 493 44. Štefanac T.; Grgas D.; Landeka Dragičević T., 2021. Xenobiotics—Division and  
494 Methods of Detection: A Review. *J. Xenobiot.*, 11, 130–141. DOI: 10.3390/jox11040009
- 495 45. Tang H., Liu Y., Liu X., Zhang A., Yang R., Han Y., Liu P., He H., Li Z., 2023. Regulation  
496 methods and enhanced mechanism on the efficient degradation of aromatics in biochemical  
497 treatment system of coal chemical wastewater. *Journal of Environmental Management*,  
498 348, 119358, DOI: 10.1016/j.jenvman.2023.119358.
- 499 46. Tutić M., Miloloža M. Cvetnić V., Martinja, L., Furač, M., Markić Š., Ukić T., Bolanča,  
500 Kučić Grgić D., 2023. An Overview of Coking Wastewater Characteristics and Treatment  
501 Technologies, *Kem. Ind.* 72 (5-6) 349–358.
- 502 47. U.S. Environmental Protection Agency, 2014 Toxic and Priority Pollutants Under the Clean  
503 Water Act | Effluent Guidelines | US EPA U.S. Washington
- 504 48. Villegas L.G.C., Mashhadi N., Chen M., Mukherjee D., Taylor K.E., Biswas N., 2016. A  
505 Short Review of Techniques for Phenol Removal from Wastewater. *Curr Pollution Rep*, 2,  
506 157–167. DOI: 10.1007/s40726-016-0035-3
- 507 49. Wu X., Zhang C., An H., Li M., Pan X., Dong F., Zheng Y., 2021. Biological removal of  
508 deltamethrin in contaminated water, soils, and vegetables by *Stenotrophomonas*  
509 *maltophilia* XQ08. *Chemosphere* 279, 130622. DOI: 10.1016/j.chemosphere.2021.130622

- 510 50. Zhang J., Zhou X., Zhou Q., Zhang J., Liang, J., 2022. A study of highly efficient phenol  
511 biodegradation by a versatile *Bacillus cereus* ZWB3 on aerobic condition. *Water Science  
512 and Technology*, 86, 355–366. DOI: 10.2166/wst.2022.209
- 513 51. Zhang J., Bing W., Hu T., Zhou X., Liang J., Li Y., 2023. Enhanced biodegradation of  
514 phenol by microbial collaboration: Resistance, metabolite utilization, and pH stabilization.  
515 *Environmental Research*, 238, Part 2, 117269, DOI: 10.1016/j.envres.2023.117269.
- 516 52. Zhao T., Gao Y., Yu T., Zhang Y., Zhang Z., Zhang I., Zhang L., 2021. Biodegradation of  
517 phenol by a highly tolerant strain *Rhodococcus ruber* C1: Biochemical characterization  
518 and comparative genome analysis. *Ecotoxicology and Environmental Safety*, 208, 111709,  
519 DOI: 10.1016/j.ecoenv.2020.111709.

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