# JOURNAL OF PLANT PROTECTION RESEARCH Vol. 47, No. 2 (2007)

# DIFFERENTIAL ANTAGONISM OF TRICHODERMA SP. AGAINST MACROPHOMINA PHASEOLINA

Aly A. Aly<sup>1</sup>, Mohamed A. Abdel-Sattar<sup>2</sup>, Moawad R. Omar, Kamel A. Abd-Elsalam<sup>1\*</sup>

<sup>1</sup>Plant Pathology Research Institute, Agricultural Research Center, 9-Gamaa, St. Giza, Egypt

Accepted: March 9, 2007

**Abstract:** In view of the ecological hazards of chemicals, pot experiments were conducted to determine the efficacy of *Trichoderma* sp. against *Macrophomina phaseolina*. Greenhouse evolution of the interaction between *M. phaseolina* isolates and *Trichoderma* sp. isolates revealed a very highly significant (p=0.0000). *M. phaseolina* isolate x antagonist isolate interaction for all the following parameters: preemergence damping-off, postemergence damping-off, survival, plant height, and dry weight. This interaction implies that a single isolate of antagonist can be highly effective against an isolate of *M. phaseolina*, but may have only minimal effects on other isolates of *M. phaseolina*. Therefore, isolates of antagonist should be tested against as many isolates of *M. phaseolina* as possible, as this will improve the chance of identifying antagonist isolates effective against several isolates of *M. phaseolina*.

Key words: cotton, Macrophomina, Trichoderma, seed coating

#### INTRODUCTION

Charcoal root rot (CRR) is an economically important disease of many crops especially in cotton and soybean. *Macrophomina phaseolina* (Tassi) Goid. a soilborne fungus causes charcoal root rot. The fungus can infect the root and lower stem of over 500 plant species (Wyllie 1989). The pathogen is a widespread distribution in the Egyptian soil, and it is easily and frequently isolated from cotton roots particularly during the late period of the growing season (Omar 1999). Although initial infections of cotton by *M. phaseolina* occur at seedling stage, they usually remain latent until the cotton plant approaches maturity (Dhingra and Sinclair 1978). Aly et al. (2006) found that resistance to *M. phaseolina* was completely lacking in the commercial Egyptian cottons (*Gossypium barabadense* L.). Thus, the use of seed-dressing fungicides for con-

<sup>&</sup>lt;sup>2</sup>Deptement of Agricultural Botany Faculty of Agriculture, Suez Canal University, Ismailia, Egypt

<sup>\*</sup>Corresponding author:

Plant Pathology Research Institute, Agricultural Research Center, 9-Gamaa, St. Giza, Egypt abdelsalamka@gmail.com

trolling the disease has become indispensable under Egyptian conditions. While effective fungicides are available (Abdel-Aziz et al. 1996; Aly et al. 2001), it is becoming increasingly evident that their widespread use is associated with some problems, such as the potential harmful effect on non-target organisms, the development of resistance races of the pathogen, and the possible carcinogenicity. The continuous use of chemical treatments has resulted in control failure when the pathogens become resistant to the active ingredient (Williams and Gisi 1992). Other problems include gradual elimination and phasing out of some compounds (Zaki et al. 1998).

Biological control has been considered as a serious alternative to seed-dressing fungicides. Regarding *M. phaseolina*, a number of reports demonstrated that some fungi, in particular *Trichoderma* spp. could be effectively used for the suppression of this pathogen (Chowdhury 1998; Khan and Gupta 1998; Raguchander et al. 1997, 1998; Singh and Sindhan 1998; Rajurkar et al. 1998; Adekunle et al. 2001; Aly et al. 2001). Other workers (Kumar and Khare 1990; Parveen and Ghaffar 1991; Mathur 2006) have also proposed T. *harziahum* as a potential biocontrol agent of *M. phaseolina*.

The objective of this study was to evaluate the interaction between 5 isolates of *Trichoderma* sp. and 14 isolates of *M. phaseolina*, pathogenic on cotton, under greenhouse conditions.

#### MATERIALS AND METHODS

### **Fungal** isolates

Isolates of *Trichoderma* sp. and *M. phaseolina* used in the current study (Tables 1, 2) were obtained from the fungal collection of Cotton Disease Research Section, Plant Pathology Research Institute, Agricultural Research Center, Giza, Egypt. *Trichoderma* sp. were originally isolated from cotton roots, while *M. phaseolina* isolates were recovered from cotton and other hosts.

Table 1. Geographic origin of *Trichoderma* spp. used in studying the interaction between isolates of *M. phaseolina* and isolates of *Trichoderma* spp.

Isolate No.	Geographic orgin
1	Sharqiya
2	Daqahliya
3	El-Minya
4	Giza
5	Assiut

# Production of M. phaseolina inoculum used in soil infestation

Substrate for growth of isolates was prepared in 500-ml glass bottles, each bottle contained 100 g of sorghum grains and 80 ml of tap water. Contents of each bottle were autoclaved for 30 minutes. Isolate inoculum, taken from one-week-old culture on PDA, was aseptically introduced into the bottle and allowed to colonize sorghum for three weeks.



Table 2.	Geographic	origin and	sources of M.	phaseolina isolates

Isolate No.	Geographic origin	Source	
1	Giza	soybean	
2	Giza	sunflower	
3	Beheira	cotton	
4	Kafr El-Sheikh	cotton	
5	Faiyoum	sesame	
6	Giza	sesame	
7	Beheira	cotton	
8	Giza	cotton	
9	Daqahliya	cotton	
10	Daqahliya	cotton	
11	Kafr El-Sheikh	cotton	
12	Giza	soybean	
13	Gharbiya	cotton	
14	Sharqiya	cotton	

# Production of Trichoderma sp. inoculum used in seed treatment

Inoculum of *Trichoderma* sp. isolates was prepared as previously mentioned; however, antagonist-sorghum mixture was air-dried in the greenhouse. The dry mixture was triturated to a fine powder in a blender (Papavizas and Lewis 1981).

#### In vivo interaction between *Trichoderma* sp. and *M. phaseolina* isolates

Fourteen batches of autoclaved clay loam soil were placed on greenhouse benches and individually infested with inoculum of each M. phaseolina isolates at the rate of  $40\,\mathrm{g/kg}$  soil. The inoculum consisted of mycelia and scleroia growing on sorghum. After thoroughly mixing, infested soil was dispensed into 15-cm-diameter clay pots. Seeds of cultivar Giza 89 were treated with the powdered inoculum of each isolate of Trichoderma sp. at the rate of  $6\,\mathrm{g/kg}$  seeds.

In the control treatment, seeds were treated with sorghum powder at the same rate. Slightly moist seeds were treated with inoculum of each isolate, and thoroughly shaken in plastic bags before being planted at the rate of 10 seeds/pot of M. phaseolina-infested soil. The pots (5 for each treatment) were randomly distributed on a greenhouse bench under a temperature regime ranging from  $19.5\pm1.5$  to  $34\pm4^{\circ}$ C. Preemergence damping-off was recorded 15 days after planting. Preemergence damping-off, survivals, plant height (cm), and dry weight (mg/plant) were recorded 45 days after planting.

#### Analysis of statistical data

The experimental design of the present study was a randomized complete block design with five replicates. Analysis of variance (ANOVA) of the data was performed with the MSTAT-C Statistical Package. Least significant difference (LSD) was used to compare between means of *Trichoderma* sp. isolates within *M. phaseolina* isolates. Percentage data were transformed into arc sine angles before carrying out the ANOVA to produce approximately constant variance.



# **RESULTS**

ANOVA (Table 3) showed very highly significant (p=0.0000) effects of *Trichoderma* isolate, *M. phaseolina* isolate, and their interaction on all the tested parameters. *Trichoderma* isolate x *M. phaseolina* isolate interaction was the most important factor in determining variation in all the tested parameters.

Table 3. Analysis of variance of *Trichoderma* isolates, *M. phaseolina* isolate and their interaction on cotton seedling disease variables (cultivar Giza 89) under greenhouse conditions

Parameter and source of variation	D.F.	M.S.	F. value	P > F
Preemergence damping-off				
Replication	4	18,011	0,4479	
Trichoderma isolate (T)	5	1102,647	27,4206	0.0000
M. phaseolina isolate (M)	13	929,316	23,1102	0.0000
$T \times M$	65	336,265	8,3622	0.0000
Error	332	40,212		
Postemergence damping-off				
Replication	4	373,663	5,3389	0.0004
Trichoderma isolate (T)	5	2522,831	36,0461	0.0000
M. phaseolina isolate (M)	13	2077,617	29,6849	0.0000
T x M	65	941,618	13,4538	0.0000
Error	332	69,989		
Survival				
Replication	4	371,363	5,7446	0.0002
Trichoderma isolate (T)	5	5770,213	89,2595	0.0000
M. phaseolina isolate (I)	13	1 648,693	25,5036	0.0000
$T \times M$	65	680,381	10,5248	0.0000
Error	332	64,645		
Plant height				
Replication	4	79,403	2,3832	0.0513
Trichoderma isolate (T)	5	744,693	22,3514	0.0000
M. phaseolina isolate (M)	13	139,608	4,1902	0.0000
$T \times M$	65	97,324	2,9211	0.0000
Error	332	33,318		
Dry weight				
Replication	4	61 562,640	2,2453	0.0639
Trichoderma isolate (T)	5	712015,734	25,9690	0.0000
M. phaseolina isolate (M)	13	82 193,485	2,9978	0.0000
T x M	65	100785,166	3,6759	0.0000
Error	332	27417,890		

<sup>&</sup>lt;sup>a</sup> replication is random, while each of *Trichoderma* isolate and *M. phaseolina* isolate is fixed

D.F. - Degrees of freedom

M.S. - mean square

F. – F. value used to test the hypothesis of equal population means P-value is the area to the right of the F statistic under an F distribution

with g-1 and N-g degrees of freedom

*Trichoderma* isolate was the second in importance as a source of variation in survival, plant height, and dry weight, while *M. phaseolina* isolate was the second in importance as a source of variation in preemergence damping-off and postemergence damping-off (Table 4).

Table 4. Relative contribution of *Trichoderma* isolate, *M. phaseolina* isolates and their interaction to variation in cotton seedling disease variables (cultivar Giza 89) under greenhouse conditions

		Relative co	ntribution <sup>a</sup> to v	variation in	
Source of variation	pre- emergence damping-off	post- emergence damping-off	survival	plant height	dry weight
Trichoderma isolate (T)	13.96	12.33	30.05	30.57	31.16
M. phaseolina isolate (M)	30.57	26.40	22.33	14.89	9.35
TxM	55.30	59.82	46.07	51.93	57.34

<sup>&</sup>lt;sup>a</sup> calculated as percentage of sum squares of the explained (model) variation

Due to the very highly significant effect of Trichoderma isolate x M. phaseolina isolate interaction on preemergence damping-off, LSD was calculated to compare means of Trichoderma isolates within each isolate of M. phaseolina (Table 5). These comparisons showed that the differences in preemergence damping-off between Trichoderma isolates and the control were not the same for each M. phaseolina isolate that is, M. phaseolina isolates responded differently to the application of Trichoderma isolates. For example, Trichoderma 1 was the only isolate, which significantly reduced preemergence damping-off caused by M. phaseolina 1. Preemergance damping-off caused by M. phaseolina 5 was significantly suppressed by all the Trichoderma isolates; however, Trichoderma isolates showed different levels of efficiency in suppressing this isolate of M. phaseolina. It is worth noting that some Trichoderma isolates proved to be stimulatory for pathogenicity of some M. phaseolina isolates like Trichoderma 3 and Trichoderma 4, which significantly increased pathogenicity of M. phaseolina 12 and M. phaseolina 10, respectively. It was also found that the magnitude of the differences between Trichoderma isolates differed from one M. phaseolina isolate to another. For example, the difference between Trichoderma 1 and Trichoderma 2 was highly significant against M. phaseolina 1, while it was nonsignificant against M. phaseolina 3. The difference between Trichoderma 4 and Trichoderma 5 was nonsignificant against M. phaseolina 13, while it was highly significant against M. phaseolina 14. The previously mentioned conclusions regarding preemergence damping-off hold true for postemergence damping-off data shown in Table 6, survival data shown in Table 7, and data of seedling growth parameters (Table 8).

www.czasopisma.pan.pl

Effect of Trichoderma isolate, M. phaseolina isolates and their interaction on preemergence damping-off of cotton seedlings (cultivar Giza 89) under greenhouse conditions Table 5.

						Is	olate of 5	Isolate of Trichoderma sp.	ф					
M. phaseolina isolate	Trich	Trichoderma 1	Trich	Trichoderma 2	Trich	Trichoderma 3	Trich	Trichoderma 4	Trich	Trichoderma 5	Ŭ	Control		Mean
	[%]	Transfor- med	[%]	Transfor- med	[%]	Transfor- med	[%]	Transfor- med	[%]	Transfor- med	[%]	Transfor- med	[%]	Transfor- med
1	14.00ª	(21.69) <sup>b</sup>	34.00	(35.02)	24.00	(28.80)	16.00	(23.02)	22.00	(27.60)	26.00	(30.55)	22.67	(27.78)
2	16.00	(20.95)	36.00	(36.82)	20.00	(26.27)	18.00	(22.28)	26.00	(30.55)	42.00	(40.33)	26.33	(29.54)
3	28.00	(31.88)	24.00	(29.09)	10.00	(16.38)	14.00	(19.33)	20.00	(26.27)	18.00	(24.35)	19.00	(24.55)
4	36.00	(36.65)	10.00	(18.44)	34.00	(35.49)	26.00	(30.55)	22.00	(27.60)	36.00	(36.65)	27.33	(30.90)
5	34.00	(35.62)	42.00	(40.33)	74.00	(47.31)	30.00	(32.96)	18.00	(24.35)	68.00	(55.59)	44.33	(36.36)
9	34.00	(35.02)	50.00	(45.00)	26.00	(48.46)	20.00	(25.85)	30.00	(32.78)	54.00	(47.31)	40.67	(39.07)
7	38.00	(37.98)	26.00	(30.55)	68.00	(55.59)	34.00	(35.49)	48.00	(43.85)	54.00	(47.31)	44.67	(41.79)
8	26.00	(30.55)	24.00	(29.22)	28.00	(31.28)	00:09	(50.77)	54.00	(47.26)	44.00	(41.54)	39.33	(38.44)
6	32.00	(34.29)	28.00	(31.75)	50.00	(45.00)	38.00	(39.13)	48.00	(43.85)	62.00	(51.97)	43.00	(40.99)
10	22.00	(27.89)	38.00	(38.03)	30.00	(32.66)	26.00	(48.46)	32.00	(34.24)	18.00	(24.64)	32.67	(34.32)
11	10.00	(16.38)	18.00	(24.64)	28.00	(31.33)	28.00	(31.88)	42.00	(40.33)	24.00	(28.80)	25.00	(28.89)
12	12.00	(20.06)	14.00	(21.69)	60.00	(48.51)	36.00	(36.65)	16.00	(23.31)	34.00	(35.27)	28.67	(30.92)
13	20.00	(26.56)	26.00	(30.42)	40.00	(39.13)	14.00	(21.69)	12.00	(20.06)	32.00	(34.29)	24.00	(28.69)
14	10.00	(16.38)	34.00	(38.03)	40.00	(38.95)	00:09	(50.87)	12.00	(20.06)	46.00	(42.64)	33.67	(34.49)
Mean	23.71	(27.99)	28.86	(32.07)	40.14	(37.51)	32.14	(33.50)	28.71	(31.58)	39.86	(38.66)	32.24	(33.55)
Cotocilore Craft to me come	1:00+00													

mean of five replicates

<sup>&</sup>lt;sup>b</sup> percentage data were transformed into arc sine angles before carrying out the analysis of variance to produce approximately constant variance LSD (transformed data) for isolate of *Trichoderma* sp. x isolates of *M. phaseolina* interaction = 7.89 ( $p \le 0.05$ ) or 10.39 ( $p \le 0.01$ )

Table 6. Effect of Trichodernna isolate, M. phaseolina isolate, and their interaction on postemergence damping-off of cotton seedlings (cultivar Giza 89) under greenhouse conditions

						Is	olate of 5	Isolate of Trichoderma sp.	ė					
M. phaseolina isolate	Trich	Trichoderma 1	Trich	Trichoderma 2	Trich	Trichoderma 3	Trich	Trichoderma 4	Trich	Trichoderma 5	Ŭ	Control	V	Mean
	[%]	Transfor- med	[%]	Transfor- med	[%]	Transfor- med	[%]	Transfor- med	[%]	Transfor- med	[%]	Transfor- med	[%]	Transfor- med
1	50.00ª	(45.05) <sup>b</sup>	10.00	(14.02)	72.00	(58.37)	10.00	(14.31)	4.00	(7.38)	52.00	(46.15)	33.00	(30.88)
2	8.00	(10.62)	8.00	(12.69)	20.00	(25.85)	12.00	(18.00)	22.00	(24.69)	34.00	(35.62)	17.33	(21.25)
3	14.00	(19.62)	50.00	(45.00)	22.00	(27.60)	26.00	(30.00)	8.00	(10.33)	52.00	(46.20)	28.67	(29.86)
4	2.00	(3.69)	40.00	(39.18)	42.00	(40.33)	42.00	(40.00)	12.00	(18.00)	34.00	(35.32)	28.67	(29.47)
5	8.00	(14.75)	40.00	(39.18)	8.00	(12.69)	40.00	(39.18)	54.00	(47.36)	28.00	(31.75)	29.67	(24.90)
9	10.00	(14.31)	2.00	(3.69)	4.00	(7.38)	20.00	(26.27)	44.00	(41.44)	40.00	(39.18)	20.00	(22.04)
7	12.00	(18.00)	12.00	(15.94)	30.00	(33.21)	10.00	(16.38)	16.00	(23.31)	34.00	(35.49)	19.00	(23.72)
8	12.00	(18.00)	10.00	(14.31)	00.9	(9.00)	4.00	(7.38)	16.00	(23.31)	38.00	(37.80)	14.33	(18.30)
6	54.00	(49.67)	52.00	(53.35)	20.00	(26.27)	14.00	(19.33)	22.00	(27.60)	34.00	(35.62)	32.67	(35.31)
10	40.00	(39.18)	34.00	(35.62)	44.00	(41.54)	26.00	(30.55)	4.00	(7.38)	74.00	(60.29)	48.33	(35.76)
11	44.00	(41.49)	50.00	(45.00)	64.00	(47.36)	00:09	(51.05)	8.00	(12.69)	64.00	(53.18)	48.00	(41.79)
12	54.00	(47.36)	50.00	(45.00)	22.00	(27.89)	46.00	(42.64)	14.00	(19.62)	00.09	(20.99)	41.00	(38.92)
13	72.00	(58.25)	30.00	(32.96)	46.00	(42.64)	00.99	(54.51)	26.00	(48.51)	48.00	(43.80)	53.00	(46.78)
14	30.00	(32.49)	10.00	(16.38)	52.00	(46.20)	4.00	(7.38)	58.00	(49.67)	42.00	(40.33)	32.67	(32.07)
Mean	29.29	(29.46)	28.43	(26.92)	32.29	(31.88)	27.14	(28.41)	23.43	(25.81)	45.29	(42.27)	30.98	(30.78)

a mean of five replicates

<sup>&</sup>lt;sup>b</sup> percentage data were transformed into arc sine angles before carrying out the analysis of variance to produce approximately constant variance LSD (transformed data) for isolate of Trichoderma sp. x M. phaseolina isolates interaction =  $10.41~(p \le 0.05)$  or  $13.71~(p \le 0.01)$ 

Table 7. Effect of Trichoderma isolate, M. phaseolina isolate and their interaction on survival of cotton seedlings (Cultivar Giza 89) under greenhouse conditions

						Is	olate of	Isolate of <i>Trichoderma</i> sp.	φ					
M. phaseolina isolate	Trick	Trichoderma 1	Trich	Trichoderma 2	Trich	Trichoderma 3	Trich	Trichoderma 4	Trick	Trichoderma 5	Ŭ	Control		Mean
	[%]	Transfor- med	[%]	Transfor- med	[%]	Transfor- med	[%]	Transfor- med	[%]	Transfor- med	[%]	Transfor- med	[%]	Transfor- med
1	36.00ª	(36.65) <sup>b</sup>	56.00	(49.16)	4.00	(7.38)	74.00	(59.87)	74.00	(59.87)	24.00	(29.22)	44.67	(40.36)
2	76.00	(80.78)	56.00	(48.51)	00.09	(50.87)	70.00	(57.04)	52.00	(46.15)	24.00	(29.22)	56.33	(48.76)
3	58.00	(49.72)	26.00	(30.42)	00.89	(55.84)	26.00	(50.82)	72.00	(58.25)	30.00	(32.96)	50.00	(46.33)
4	62.00	(52.20)	50.00	(45.00)	24.00	(29.22)	32.00	(34.29)	00.99	(54.38)	30.00	(32.96)	44.00	(41.34)
5	58.00	(49.67)	18.00	(24.64)	38.00	(37.98)	30.00	(32.96)	28.00	(28.80)	40.00	(7.38)	35.33	(30.24)
9	56.00	(48.56)	48.00	(43.85)	40.00	(39.18)	00.09	(50.92)	26.00	(30.42)	00.9	(9.00)	39.33	(36.99)
7	50.00	(45.00)	62.00	(52.02)	2.00	(3.69)	56.00	(48.56)	28.00	(36.82)	12.00	(15.64)	35.00	(33.62)
8	62.00	(52.15)	66.00	(54.51)	00.99	(55.03)	36.00	(36.82)	30.00	(32.73)	18.00	(24.64)	46.33	(42.65)
6	10.00	(16.38)	8.00	(10.62)	30.00	(32.91)	46.00	(42.69)	30.00	(32.96)	4.00	(7.38)	21.33	(23.82)
10	38.00	(37.98)	28.00	(31.75)	28.00	(30.42)	18.00	(24.64)	64.00	(53.40)	8.00	(12.69)	30.67	(31.82)
11	46.00	(42.69)	32.00	(34.29)	18.00	(24.64)	12.00	(15.64)	50.00	(45.00)	12.00	(15.64)	30.00	(29.65)
12	34.00	(35.49)	36.00	(36.60)	22.00	(25.11)	18.00	(24.64)	70.00	(57.04)	00.9	(9.00)	31.00	(31.31)
13	8.00	(12.69)	44.00	(41.54)	14.00	(21.69)	20.00	(26.27)	32.00	(34.29)	20.00	(26.27)	23.00	(27.12)
14	60.00	(51.05)	52.00	(46.15)	8.00	(12.69)	36.00	(36.82)	28.00	(31.75)	16.00	(18.60)	33.33	(32.84)
Mean	46.71	(42.21)	41.57	(39.22)	30.14	(30.47)	40.29	(38.71)	46.43	(42.99)	17.86	(19.33)	44.31	(35.49)

a mean of five replicates

<sup>\*</sup> percentage data were transformed into arc sine angles, before carrying out the analysis of variance to produce approximately constant variance LSD (transformed data) for isolate of Trichoderma  $\times$  M. phaseolina isolates interaction = 10.00 (p  $\leq$  0.05) or 13.17 (p  $\leq$  0.01)



Table 8. Effect of *Trichoderma* isolates, *M. phaseolina* isolates, and their interaction on plant height and dry weight of cotton seedlings (cultivar Giza 89) under greenhouse conditions

M.				Plant heig	ht [cm]		
phase- olina isolate	Tricho- derma 1	Tricho- derma 2	Tricho- derma 3	Tricho- derma 4	Tricho- derma 5	control	mean
1	22.86a	24.90	9.85	24.88	22.81	20.10	20.90
2	27.27	26.32	22.29	27.24	20.77	18.60	23.75
3	26.17	29.36	23.07	25.86	24.85	20.85	25.03
4	22.16	22.73	25.89	27.88	23.87	22.28	24.13
5	23.82	20.56	21.09	29.57	19.46	10.50	20.84
6	23.47	29.53	20.84	20.52	20.76	10.72	20.97
7	22.51	19.80	5.70	33.19	21.18	13.01	19.23
8	28.95	23.39	20.31	27.11	22.91	21.65	24.05
9	17.61	10.44	21.90	23.43	21.57	9.75	17.45
10	22.98	21.21	25.20	24.00	23.96	14.33	21.95
11	23.95	25.02	26.07	12.55	24.01	14.95	21.09
12	23.91	22.11	20.52	23.81	20.40	9.24	20.00
13	14.10	19.91	22.09	24.13	25.26	21.46	21.16
14	23.64	25.85	12.51	24.80	17.44	12.35	19.43
Mean	23.10	22.94	19.81	24.93	22.09	15.70	

<sup>&</sup>lt;sup>a</sup> mean of five replicates

LSD for isolate of *Trichoderma* sp. x isolate of *M. phaseolina* interaction = 7.18 ( $p \le 0.05$ ), 9.46 ( $p \le 0.01$ )

М.			D	ry weight [	mg/plant]		
phase- olina isolate	Tricho- derma 1	Tricho- derma 2	Tricho- derma 3	Tricho- derma 4	Tricho- derma 5	control	mean
1	598.40	665.40	303.60	766.00	632.20	516.20	580.33
2	604.40	510.00	343.20	673.80	542.60	536.40	535.07
3	339.00	641.80	515.80	679.20	618.80	516.00	551.77
4	573.80	551.00	608.80	796.60	682.80	431.20	607.37
5	671.40	651.80	647.40	704.20	531.80	295.80	583.73
6	704.00	394.80	741.60	636.80	662.60	283.20	570.50
7	333.20	257.80	164.00	812.80	709.80	382.80	443.40
8	720.60	432.40	612.20	757.40	640.80	455.60	603.17
9	676.00	298.60	566.00	742.80	751.60	221.00	542.67
10	575.80	482.40	240.60	598.20	587.80	409.20	482.33
11	697.80	825.40	620.80	499.40	556.20	399.20	599.80
12	618.40	765.20	480.40	581.60	849.80	264.40	593.30
13	327.20	724.60	623.80	825.40	399.20	511.40	568.60
14	500.40	616.00	502.00	587.20	349.60	273.20	471.40
Mean	567.17	558.37	497.87	690.10	608.26	392.56	

LSD for isolate of Trichoderma sp. x isolate of M. phaseolina interaction =  $206.00 (p \le 0.05)$ ,  $271.30 (p \le 0.01)$ 



# DISCUSSION

Five isolates of *Trichoderma* sp. were evaluated in vivo, to assess their antagonistic potential against Macrophomina phaseolina causing CRR of cotton. Greenhouse evaluation of the interaction between M. phaseolina isolates and Trichoderma isolates revealed a very highly significant (p = 0.0000) M. phaseolina isolate x antagonist isolate interaction for all the tested parameters. This interaction implies that a single isolate of antagonist can be highly effective against an isolate of M. phaseolina, but may have only minimal effects on the other isolates of M. phaseolina. The interaction also indicates that apparently many genes from both organisms interact to regulate the amount of antagonism between M. phaseolina and Trichoderma isolates (Wells and Bell 1983). Alv et al. (2001) reported similar interaction when they studied the in vitro antagonism of Trichoderma spp., Penicillium spp., and Aspergillus spp. against M. phaseolina isolates. The results of Cardona and Rodriguez (2006) show that there was no any effect of T. harzianum on the incidence of the charcoal rot disease in sesame. Isolates of T. koningii and T. harzianum were selected from soil dilutions and tested in vitro for their antagonistic behaviour against cowpea pathogen M. phaseolina before use in the field (Adekunle et al. 2006). These findings have an important bearing on antagonism testing methods. Isolates of antagonists should be tested against as many isolates of M. phaseolina as possible, as this will improve the chance of identifying antagonist isolates effective against several isolates of M. phaseolina. The interaction also suggests that it may be more prudent to evaluate blends of antagonist isolates for wider application against more isolates of M. phaseolina. In this investigation, the interaction between M. phaseolina isolates and the antagonist isolate was evaluated under greenhouse conditions in a soil and at temperatures favourable for the growth of both M. phaseolina and the antagonist. Under field conditions, soil nutrients and temperatures during the different periods of cotton growing season may be more favourable for M. phaseolina isolates or the antagonist isolates. Thus, the results of this work are not expected to be necessarily related to the degree of biological control that may be observed in the field, but should reflect the capacities and genetic variability of the antagonist isolates and of the various M. phaseolina isolates to resist antagonism (Bell et al. 1982).

# **CONCLUSIONS**

Biological control of cotton charcoal root rot using antagonistic fungi (Trichoderma sp.) was evaluated. It is worth noting that some Trichoderma isolates proved to be stimulatory for pathogenicity of some M. phaseolina isolates. This result is in agreement with that of Khan and Gupta (1998) who demonstrated that T. polysporum was stimulatory for radial growth of M. phaseolina on PDA. In contrary, T. viride and T. harzianum were the most effective in reducing the mycelial growth and sclerotial formation of M. phaseolina. Culture filtrates of T. viride inhibited the growth of the pathogen as well as sclerotial germination to a greater extent (Karthikeyan et al. 2006). Differences in the antagonistic performance of the pathogens were observed depending on the isolates with which they interacted. Further investigations are needed on pathogen-antagonist interactions in the complex soil ecosystem to select Trichoderma isolates, which could be utilized in field to manage soilborne plant pathogens.



# **REFERENCES**

- Abdel-Aziz M.A., Moustafa-Mahmoud S.M., Ismail A.A. 1996. Impact of Imidacloprid insecticide on efficiency of some fungicides in controlling damping-off and root rot diseases of cotton seed-lings. J. Agric. Res. Tanta Univ. 22: 243–255.
- Adekunle A.T., Cardwell K.F., Florini D.A., Ikotun T. 2001. Seed treatment with *Trichoderma* species for control of damping-off of cowpea caused by *Macrophomina phaseolina*. Biocontrol Sci. Technol. 11: 449–457.
- Adekunle A.T., Ikotun T., Florini D.A., Cardwell K.F. 2006. Field evaluation of selected formulations of *Trichoderma* species as seed treatment to control damping-off of cowpea caused by *Macrophomina phaseolina*. Afr. J. Biotechnol. 5: 419–424.
- Aly A.A., El-Shazly A.M.M., Youssef R.M., Omar M.R. 2001. Chemical and biological control of charcoal rot of cotton caused by *Macrophomina phaseolina*. J. Agric. Sci. Mansoura Univ. 26: 7661–7674.
- Aly A.A., Abdel-Sattar M.A., Omar M. R. 2006. Susceptibility of some Egyptian cotton cultivars to charcoal rot disease caused by *Macrophomina phaseolina*. J. Agric. Sci. Mansoura Univ. 31: 5025–5037.
- Bell D.K., Wells H.D., Markham C.R. 1982. In vitro antagonism of *Trichoderma* species against six fungal plant pathogens. Phytopathology 72: 379–382.
- Cardona R., Rodriguez H. 2006. Effects of *Trichoderma harzianum* fungus on the incidence of the charcoal rot disease on sesame. Rev. Fac. Agron. 23: 44–50.
- Chowdhury A.K. 1998. Biocontrol of Macrophomina infection of jute. Environ. Ecol. 16: 44-45.
- Dhingra O.D., Sinclair J.B. 1978. Biology and Pathology of *Macrophomina phaseolina*. Imprensia Universidade Federal de Viscosa, Brazil, 166 pp.
- Karthikeyan V., Sankaralingam A., Nakkeeran S. 2006. Management of groundnut root rot with biocontrol agents and organic amendments. Arch. Phytopatol. Plant Prot. 39: 215–223.
- Khan M.R., Gupta J. 1998. Antagonistic efficacy of *Trichoderma* species against *Macrophomina phaseolina* on eggplant. J. Plant Dis. Prot. 105: 387–393.
- Kumar S.M., Khare M.N. 1990. Studies on the antagonistic relationship of soybean spermosphere microflora with *Rhizoctonia bataticola* and *Sclerotium rolfsii*. J. Biol. Control 4: 72–74.
- Mathur A.C. 2006. Management of charcoal rot of cowpea caused by *Macrophomina phaseolina* by using biocontrol agents. Res. Crops 7: 558–560.
- Omar M.R. 1999. Studies on susceptibility of cotton to *Macrophomina phaseolina*. M.Sc. Thesis, Al-Azhar Univ. Cairo, 139 pp.
- Papavizas G.C., Lewis J.A. 1981. Introduction and augmentation of microbial antagonists for the control of soil borne plant pathogens. p. 305–322. In: "Biological Control in Crop Production (BARC Symposium No.5) (Gorge C. Papavizas, ed.). Allahnheld, Osmun, Totowa.
- Parveen S., Ghaffar A. 1991. Effect of microbial antagonists in the control of root rot of tomato. Pak. J. Bot. 23:179–182.
- Raguchander T., Rajappan K., Samiyappan R. 1997. Evaluating methods of application of biocontrol agent in the control of mungbean root rot. Indian Phytopathol. 50: 229–234.
- Raguchander T., Rajappan K., Samiyappan R. 1998. Influence of biocontrol agents and organic amendments on soybean root rot. Int. J. Trop. Agric. 16: 247–252.
- Rajurkar R.B., Gade R.M., Paslawar A.N., Chauke R.P. 1998. Management of betel vine wilt through cultural and biological methods. J. Soils Crops 8: 176–178.
- Singh R., Sindhan G.S. 1998. Effect of fungicides on the incidence of dry root rot and biochemical status of chickpea plants. Plant Dis. Res. 13: 14–17.



- Wells H.D., Bell D.K. 1983. Antagonism in vitro between isolates of *Trichoderma harzianum* and *Rhizoctonia solani* AG4. Phytopathology 73, p. 507 (Abstract).
- Williams R.J., Gisi U. 1992. Monitoring pathogen sensitivity to phenylamide fungicides: Principles and Interpretation. EPPO Bull. 22: 297–322.
- Wyllie T.D., 1989. Charcoal rot. p. 30–33. In: "Compendium of Soybean Diseases" (J.B. Sinclair, P.A. Backman, eds.). The APS Press, St. Paul.
- Zaki K., Misagi I.J., Heydari A., Shatala M.N. 1998. Control of cotton seedling damping-off in the field by *Burkholderia (Pseudomunas) cepacia*. Plant Dis. 82: 291–293.

#### **POLISH SUMMARY**

# ZRÓŻNICOWANY ANTAGONIZM TRICHODERMA SP. WOBEC MACROPHOMINA PHASEOLINA

Doświadczenia szklarniowe nad współdziałaniem pomiędzy izolatami *M. phasedina* i izolatami *Trichoderma* sp. wykazały bardzo wysoką istotność (p = 0,0000) współdziałania izolatu *M. phaseolina* z isolatem grzyba antagonistycznego dla wszystkich nastepujących parametrów: zgorzel przedwschodowa, zgorzel powschodowa, przeżywalność, wysokość roślin i sucha masa. To współdziałanie sugeruje, że pojedynczy izolat grzyba antagonistycznego może być wysoce efktywny wobec izolatu *M. phaseolina*, ale może on mieć tylko minimalny efekt w stosunku do innych izolatów *M. phaseolina*. Właściwości izolatów antagonistycznych powinny więc być przebadane przy wykorzystaniu możliwie dużej liczby izolatów *M. phaseolina*, ponieważ to rozszerzy możliwość zidentyfikowania ich efektywności wobec pewnej liczby izolatów tego patogena.